

Tracking nanomotors for *in vivo* applications using nuclear imaging techniques

Cristina Simó^a, Tania Patiño^b, Ana Hortelano^b, Unai Cossío^a, Vanessa Gómez-Vallejo^a, Jordi Llop^a, Samuel Sánchez^{b,c}

^aCIC BiomaGUNE, Paseo Miramón 182, 20014 San Sebastián, Guipúzcoa, Spain

^bInstitute for Bioengineering of Catalonia (IBEC), Baldiri Reixac 10-12, 08028 Barcelona, Spain

^cInstitució Catalana de Recerca i Estudis Avançats (ICREA), Passeig de Luís Companys, 23, 08010 Barcelona, Spain
csimo@cicbiomagune.es

Micro/nanomotors, which are micro/nanoscale devices capable to induce self-propulsion in fluid environments, have recently gained attention for different biomedical applications [1,2]. Initial works reported in the literature described micro/nanomotors able to swim in non-biocompatible matrixes such as aqueous hydrogen peroxide. Moving towards *in vivo* applications, enzyme catalysis has emerged as a biocompatible and powerful alternative to produce self-propulsion. However, tracking these micro- and nano-devices *in vivo* and at the whole body level still remains a challenge. Here, we describe the application of radiolabelling followed by Positron Emission Tomography (PET) imaging to achieve the time-resolved tracking of urease-functionalized nanomotors as potential drug delivery agents for bladder cancer.

Fully mesoporous silica nanoparticles (MSNP) with a mean diameter of 480 nm were fabricated and modified with amine groups using previously reported methods [3]. The MSNP were then functionalized with urease and heterobifunctional H₂N-PEG-SH via glutaraldehyde crosslinking, to obtain pegylated urease-powered nanomotors. The available thiol groups were then used for anchoring 20 nm gold nanoparticles. The efficient radio-fluorination of the nanomotors was achieved by the reaction (pH = 8, room temperature, 35 min) between the amine groups of the urease and 6-[¹⁸F]fluoronicotinyl-2,3,5,6-tetrafluorophenyl ester ([¹⁸F]FPy-TFP), which was prepared following a previously reported procedure [4]. PET Imaging studies in a labyrinth-type phantom (Fig. 1a) clearly showed the self-propelling capacity of the nanomotors in the presence of urea (Fig. 1c) while no significant movement was observed in pure water (Fig. 1b). *In vivo* biodistribution studies in rodents after intravenous administration showed accumulation of the nanomotors in lungs and liver, majoritarily. Our results clearly demonstrate the suitability of our approach for the *in vivo* tracking of nanomotors.

References

- [1] Vilela D., Cossío U., Parmar J., et al., *ACS Nano*, **2018**, *12*, 1220-1227.
- [2] Patiño T., Arqué X., Mestre R., et al., *Acc. Chem. Res.* **2018**, *51*, 2662-2671.
- [3] Hortelao A. C., Carrascosa R., Murillo-Cremaes N., et al., *ACS Nano*, **2019**, *13*, 429-439.
- [4] Olberg D. E., Arukwe J. M., Grace D., et al., *J. Med. Chem.* **2010**, *53*, 1732-1740.

Figures

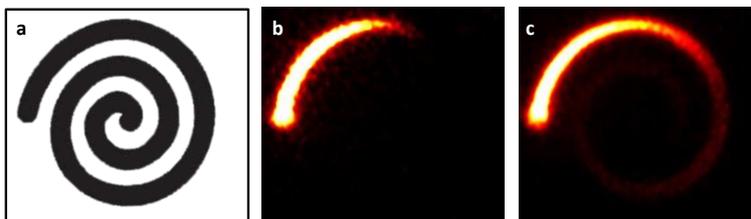


Figure 1: (a) Schematic top view of the labyrinth-type phantom; (b) Image obtained at t=60 min after seeding of nanomotors in water; (c) Image obtained at t=60 min after seeding of nanomotors in 300 mM aqueous urea solution.