# Imaging microviscosity in mesenchymal stem cells and their differentiated counterparts using a viscosity-sensitive molecular nanorotor and FLIM

## Džiugas Jurgutis<sup>1,2</sup>

Greta Jarockytė<sup>1</sup>, Aurimas Vyšniauskas<sup>2</sup>, Vitalijus Karabanovas<sup>1,2,3</sup>, Ričardas Rotomskis<sup>1,4</sup>

<sup>1</sup>Biomedical Physics Laboratory, National Cancer Institute, P. Baublio str. 3b, Vilnius, Lithuania

<sup>2</sup>State research institute Center for Physical Sciences and Technology, Saulėtekio av. 3, Vilnius, Lithuania

<sup>3</sup>Department of Chemistry and Bioengineering, Vilnius Gediminas Technical University, Saulėtekio av. 9-III, Vilnius, Lithuania

<sup>4</sup>Biophotonics group of Laser Research Centre, Vilnius University, Saulėtekio av. 9, Vilnius, Lithuania

### dziugas.jurgutis@nvi.lt

Changes microscopic in viscosity (microviscosity) signal about the ongoing intracellular alterations or the onset of various diseases, diabetes e.g. or atherosclerosis [1]. Imaging microviscosity before and after differentiation would enable a areater understanding of the differentiation mechanism and a more accurate characterization of stem cells, e.g. mesenchymal stem cells (MSC), which are able to differentiate into fat, bone or other connective tissue. Microviscosity bioimaging can be achieved by utilising molecular nanorotors with fluorescence lifetime imaging microscopy (FLIM) [1,3]. In this work, we utilise BDP-H molecular nanorotor (Figure 1). In low-viscosity environments, intramolecular rotation of BDP-H mesophenyl ring is unhindered, and the relaxation from the excited state primarily occurs through a non-radiative energy transition leading to a shorter fluorescence (FL) lifetime. In more viscous environments, the rotation is restricted and energy is released through a radiative decay, thus resulting in longer FL lifetime [1,2]. The aim of our study was to determine intracellular localization of BDP-H and to apply the molecular nanorotor for microviscosity determination in live human skin MSC and their differentiated

cultures - adipocytes and osteoblasts. Live cells were imaged with a confocal laser scanning microscope. FL lifetimes of BDP-H in live MSC were determined using timecorrelated single photon counting based FLIM. Localisation experiments showed that BDP-H accumulates in lipid droplets (LD) and cytosol of live MSC (Figure 1A). In addition, BDP-H intensely accumulates in enlarged LD of adipocytes, while in osteoblasts, cytosolic staining is more pronounced. Finally, by using a calibration curve obtained in toluene-castor oil mixtures of known bulk viscosities, and the T1 of BDP-H decays obtained with FLIM (Figure 1B), we were able to assign microviscosities values of ca. 120 cP and 104 cP to LD of undifferentiated MSC and mature adipocytes, respectively.

#### References

- [1] Kuimova, M.K., Physical Chemistry Chemical Physics, 14 (2012) 12671-12686.
- [2] Toliautas, S. et al., Chemistry A European Journal, 25 (2019) 10342-10349.
- [3] Vyšniauskas, A. & Kuimova, M.K., International Reviews in Physical Chemistry, 37 (2018) 259-285.

#### Figures



**Figure 1:** Structure of the BDP-H molecular nanorotor and its accumulation in lipid droplets of MSC (A). B – FLIM image obtained by applying a biexponential fit ( $\tau_1 = 871\pm3$  ps, mean lifetime – 1215±4 ps) ( $\lambda_{ex} = 490\pm5$  nm).

Acknowledgements: supported by the Research Council of Lithuania, Grant No. S-MIP-19-6.