



agefpi

Nanoparticles from biological source and polymer nanocomposites

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Biobased (Green) Materials

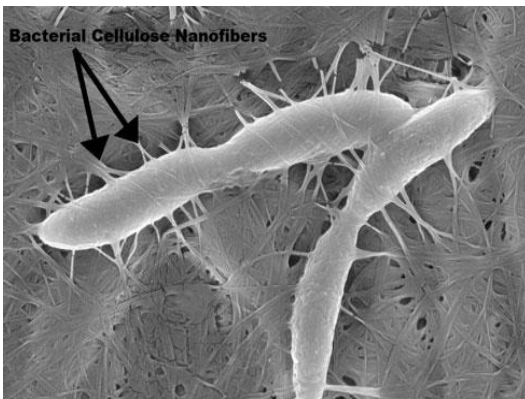
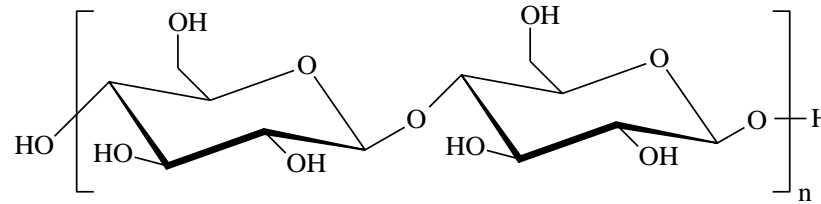
Increasing demand for products made from renewable and sustainable non-petroleum based resources (**green materials**)



Limitation: economically viable materials → **Polysaccharides?**

Cellulose

Structural material in plants, animals, bacteria



Natural (Lignocellulosic) Fibers

Reinforcing element in composites



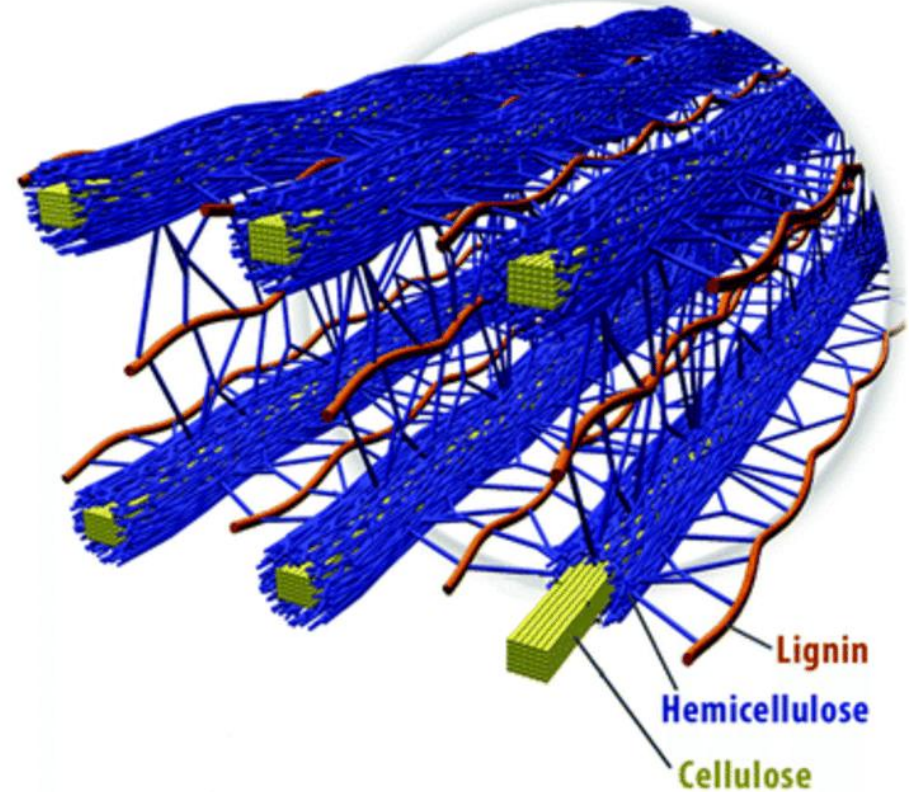
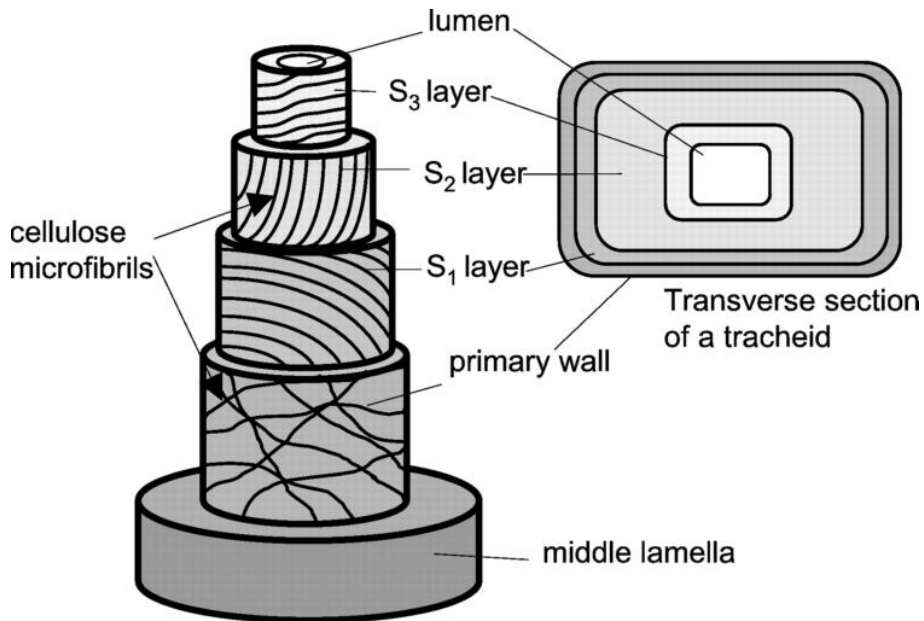
Low density, low cost, high specific strength and modulus, renewability, biodegradability, availability in a variety of forms throughout the World, flexibility, non abrasive nature to processing equipment, non-toxicity, easiness to handle, high ability for surface modification, possibility to generate energy, without residue after burning at the end of their life-cycle, economic development opportunity for non-food farm products in rural areas



Hydrophilic character : poor adhesion and dispersion in non-polar matrix, high moisture absorption, **limited thermal stability** : low permissible temperatures of processing and use

Natural (Lignocellulosic) Fibers

Structure of the cell wall



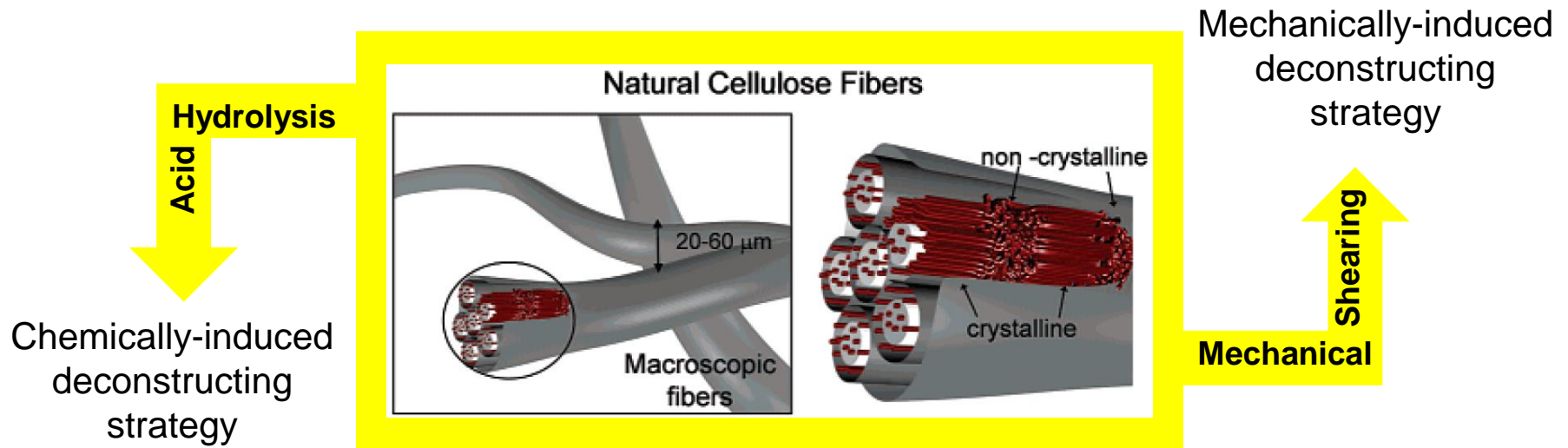
Cellulose = structural material that confers its mechanical properties to higher plant cells

Natural (Lignocellulosic) Fibers

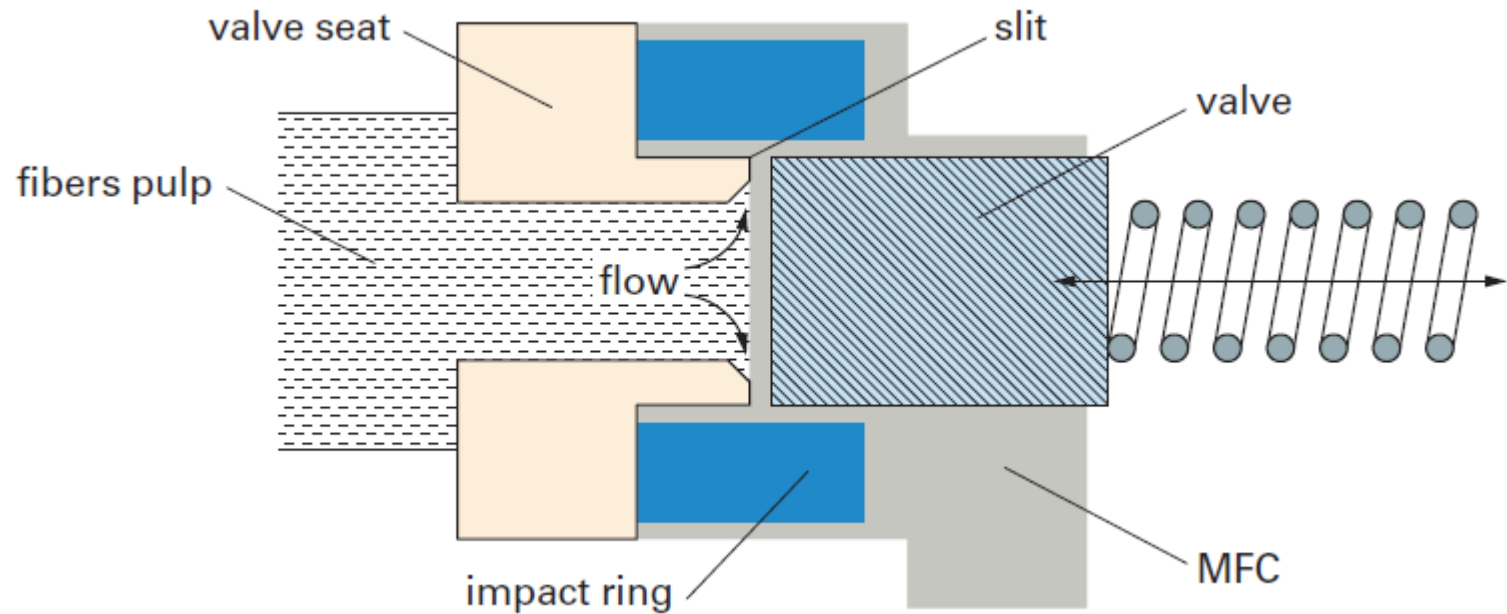
Big variation of properties inherent to the natural products
(climatic conditions, maturity, type of soil,...)
→ enormous scatter of mechanical plant fiber properties

Basic idea to achieve further improved fiber and composite is to eliminate the macroscopic flaws by disintegrating the natural grown fibers, and separating the almost defect free highly crystalline fibrils

Top-Down Deconstructing Strategy



Mechanically-induced Deconstructing Strategy



Scheme of the homogenizer

Pretreatments

High energy demand

30,000 kWh/ton (Nakagaito and Yano, 2004)

70,000 kWh/ton (Eriksen et al, 2008)

→ **necessity of a pretreatment**

Enzymatic hydrolysis

Carboxymethylation

TEMPO-catalyzed oxidation pretreatment

Cryocrushing

Mechanically-induced Deconstructing Strategy



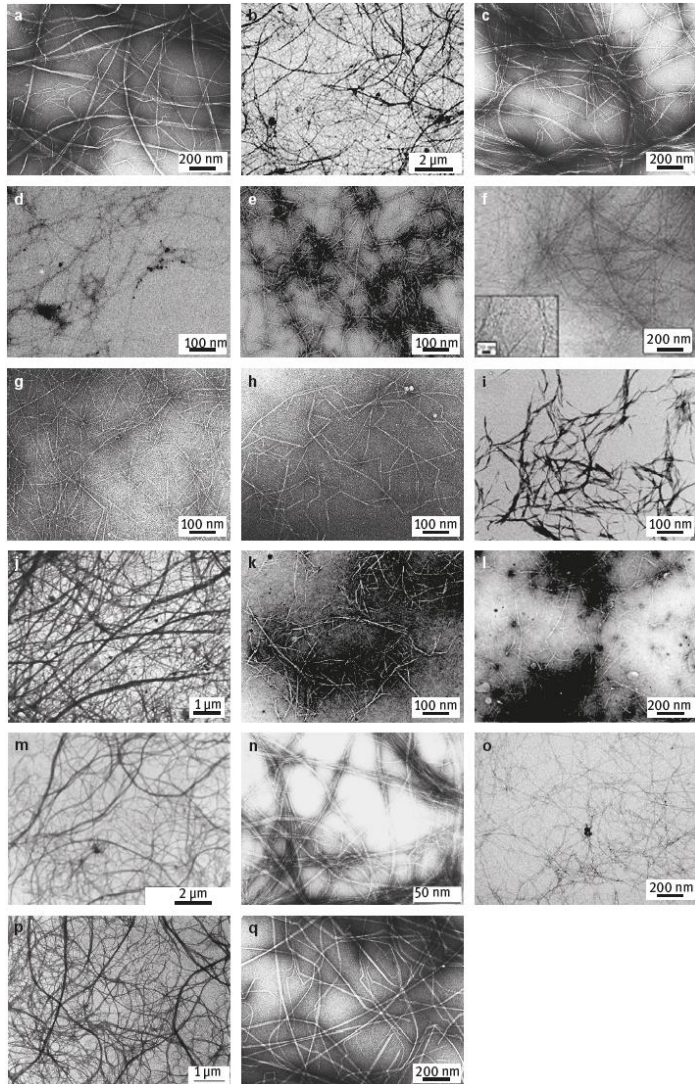
Lavoine et al., *Carbohydr. Polym.* **2012**, 90, 735-764

Width = 3-100 nm
Length > 1 μm ?



Malainine et al., *Compos. Sci. Technol.* **2005**, 65, 1520-1526

Mechanically-induced Deconstructing Strategy



TEMs showing cellulose fibers after high-pressure mechanical treatment

- (a) bacterial cellulose (Saito et al., 2006)
- (b) banana peel (Pelissari et al., 2014)
- (c) banana rachis (Zuluaga et al., 2009)
- (d) beavertail cactus (*Opuntia basilaris*) (Kakroodi et al., 2015)
- (e) bleached eucalyptus kraft pulp (Qing et al., 2013)
- (f) bleached sulfite softwood cellulose pulp (Pääkkö et al., 2007)
- (g) bleached sulfite wood pulp (Saito et al., 2006)
- (h) cotton (Saito et al., 2006)
- (i) garlic skin (Zhao et al., 2014)
- (j) *Opuntia ficus-indica* (Malainine et al., 2003)
- (k) *Posidonia oceanica* balls (Bettaieb et al., 2015)
- (l) *Posidonia oceanica* leaves (Bettaieb et al., 2015)
- (m) potato pulp (Dufresne et al., 2000)
- (n) prickly pear skin (Habibi et al., 2009)
- (o) spinifex grass (*Triodia pungens*) (Amiralian et al., 2015)
- (p) sugar beet pulp (Dufresne et al., 1997)
- (q) tunicin (Saito et al., 2006)

Dufresne, Nanocellulose: From Nature to High-Performance Tailored Materials, 2nd Ed., de Gruyter, 2017

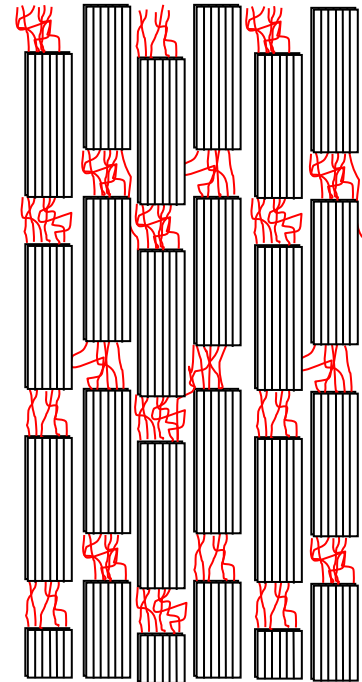
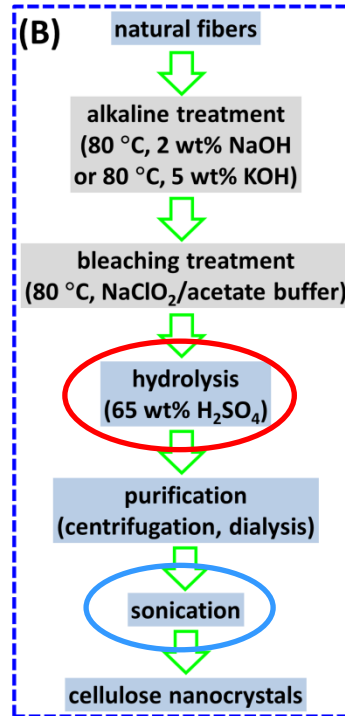
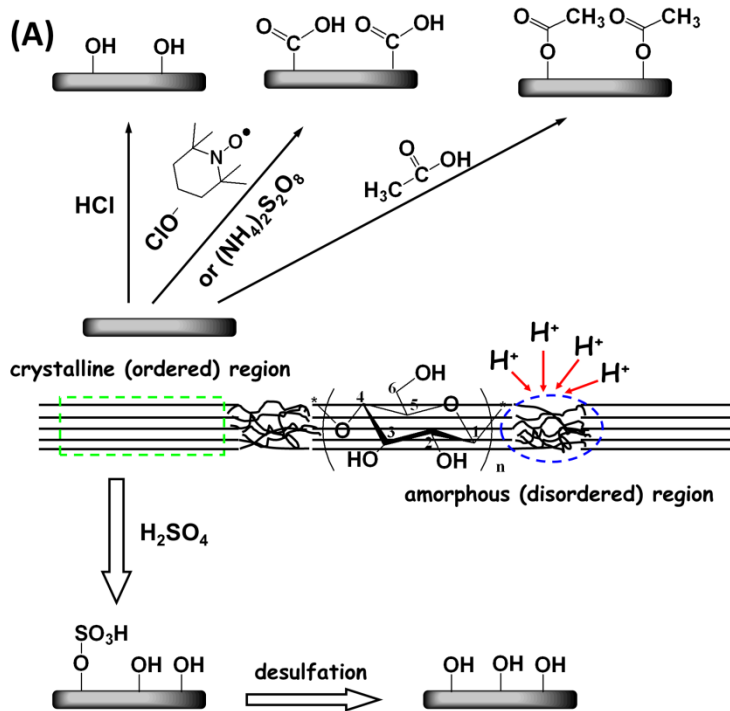
The Need for International Standards - Terminology

| Acronym | Terminology | Reference |
|---------|---------------------------------------|--|
| MFC | Microfibrillated Cellulose | (Herrick et al., 1983; Turbak et al., 1983) |
| - | Cellulose Microfibrils | (Dufresne et al., 1997; Dinand et al., 1999) |
| - | Fibrillated Cellulose | (Azizi Samir et al., 2004) |
| - | Nanofibrillar Cellulose | (Jin et al., 2004) |
| - | Fibril Aggregates | (Cheng et al., 2007) |
| - | Nanoscale Cellulose Fibrils | (Pääkkö et al., 2007) |
| - | Microfibrillated Cellulose Nanofibers | (Henriksson et al., 2007) |
| - | Cellulose Fibril Aggregates | (Cheng et al., 2007) |
| - | Cellulose Nanofibers | (Abe et al., 2007; Alemdar and Sain, 2008) |
| - | Cellulose Nanofibrils | (Henriksson et al., 2008; Ahola et al., 2008a; 2008b) |
| - | Cellulose Microfibers | (Bhattacharya et al., 2008) |
| - | Microfibril Aggregates | (Abe et al., 2009) |
| - | Cellulose Microfibril Aggregates | (Abe and Yano, 2009) |
| - | Cellulose Fibrils | (Cheng et al., 2009a; 2009b)) |
| NFC | Nanofibrillated Cellulose | (Mörseburg and Chinga-Carrasco, 2009; Chinga-Carrasco and Syverud, 2010) |
| - | Microfibrillar Cellulose | (Spence et al., 2010) |

TAPPI, ISO and
CSA approved

Table 2.1: Different terminologies used in the literature to describe the material resulting from the cellulose fiber fibrillation process.

Chemically-induced Deconstructing Strategy



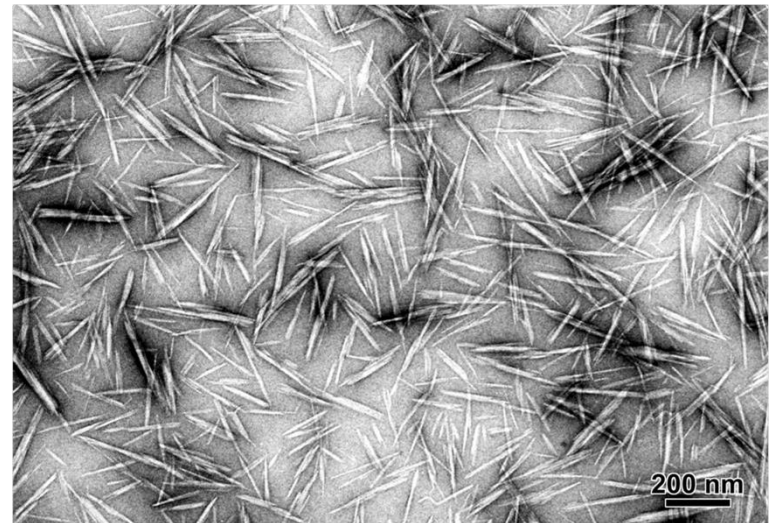
Chemically-induced Deconstructing Strategy



Siqueira et al., *Cellulose* 2010, 17, 289-298



Width = few nm
Length = few 100 nm

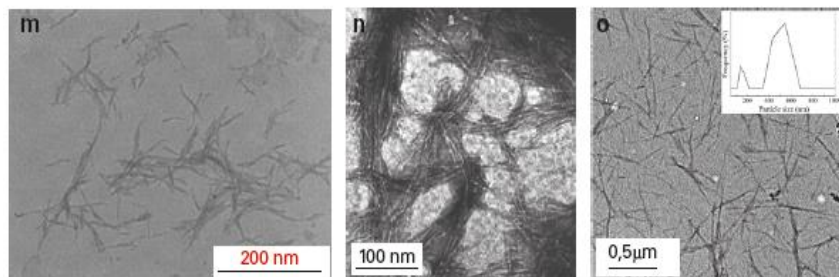
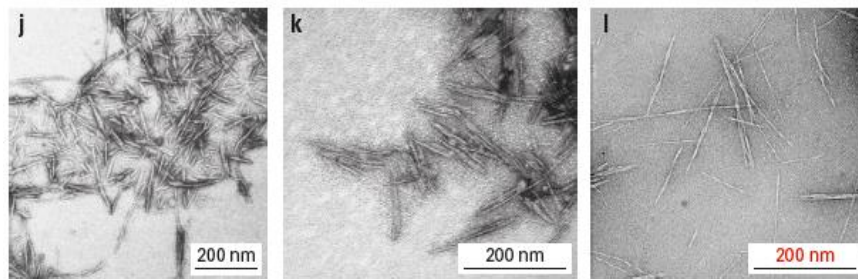
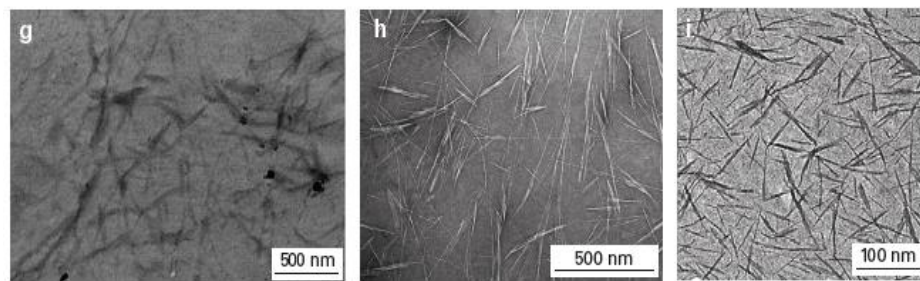
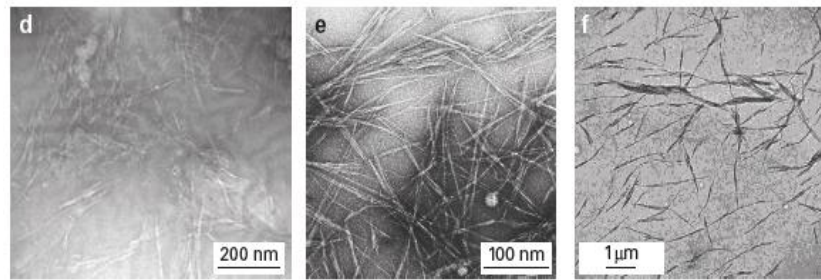
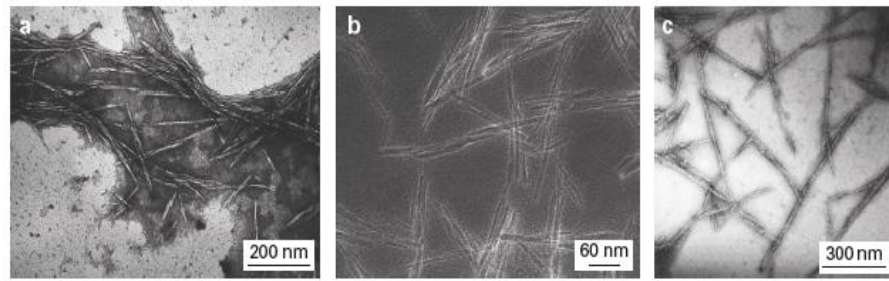


Habibi et al., *J. Mater. Chem.* 2008, 18, 5002-5010

Chemically-induced Deconstructing Strategy

TEMs from a dilute suspension of CNC from:

- (a) acacia pulp (Pu et al., 2007)
- (b) alfa (Ben Elmabrouk et al., 2009)
- (c) bacterial cellulose (Grunert and Winter, 2002)
- (d) balsa wood (Morelli et al., 2012)
- (e) banana rachis (Zuluaga et al., 2007)
- (f) bleached softwood kraft pulp (Araki et al., 1998)
- (g) brewer's spent grains (Martínez-Sanz et al., 2015)
- (h) Capim dourado (Siqueira et al., 2010)
- (i) cotton (Fleming et al., 2000)
- (j) curaúa (Corrêa et al., 2010)
- (k) eucalyptus wood pulp (de Mesquita et al., 2010)
- (l) garlic straw (Kallel et al., 2016)
- (m) giant cane (*Arundo donax*) (Barana et al., 2016)
- (n) grass of Korea (Pandey et al., 2008)
- (o) kelp residue (Feng et al., 2015) (inset: particle size distribution)



Chemically-induced Deconstructing Strategy

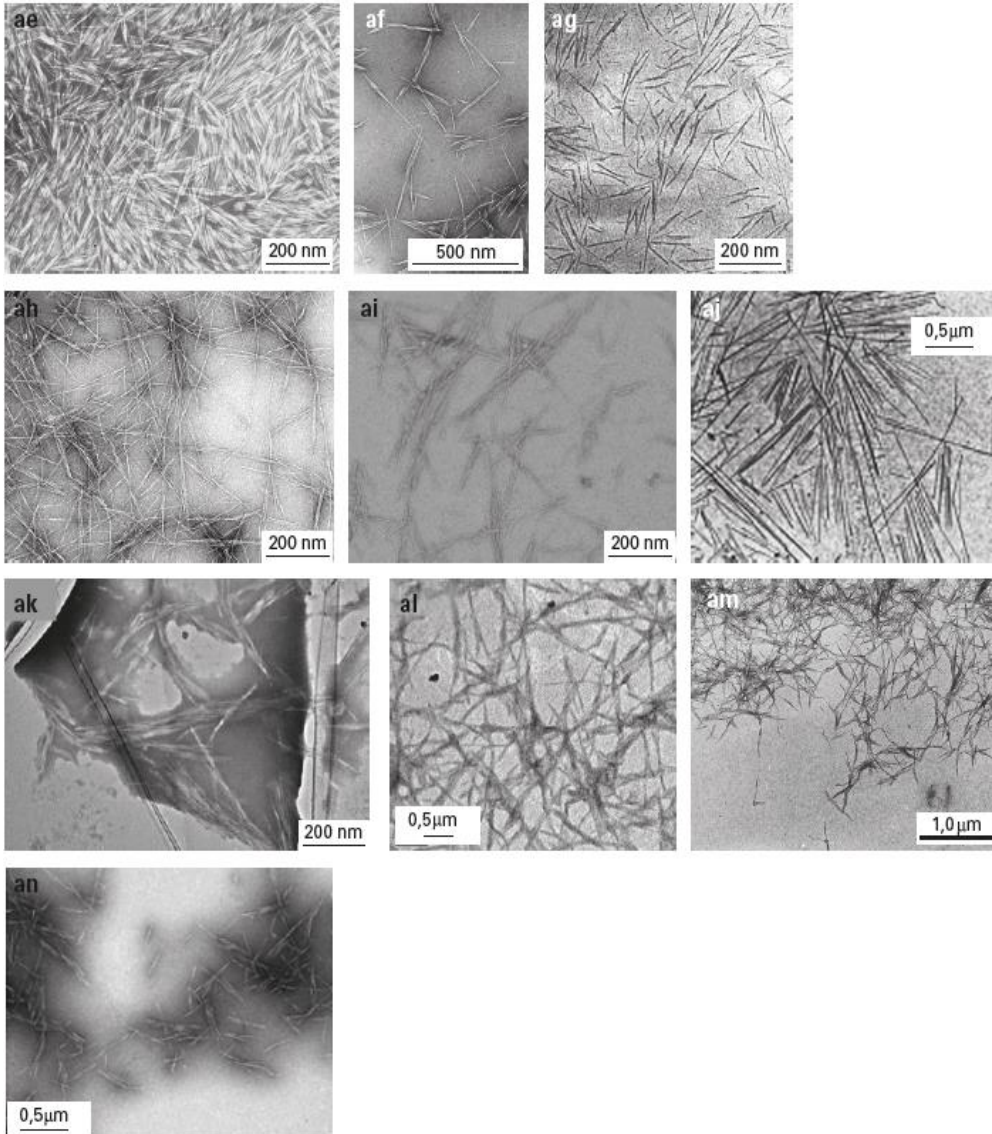
TEMs from a dilute suspension of CNC from:

- (p) kenaf (Kargarzadeh et al., 2012)
- (q) *Luffa cylindrica* (Siqueira et al., 2010)
- (r) maize straw (Rehman et al., 2014)
- (s) mango seed (Henrique et al., 2013)
- (t) MCC (Bondeson et al., 2006)
- (u) mengkuang Leaves (Sheltami et al., 2012)
- (v) oil palm trunk (Lamaming et al., 2015)
- (w) olive pomace (Martínez-Sanz et al., 2015)
- (x) olive stone (Abou-Zeid et al., 2015)
- (y) onion skin (Rhim et al., 2015)
- (z) *Pennisetum sinense* (Lu et al., 2014)
- (aa) *Posidonia oceanica* balls (Bettaieb et al., 2015)
- (ab) *Posidonia oceanica* leaves (Bettaieb et al., 2015)
- (ac) ramie (Habibi et al., 2008)
- (ad) red algae *Gelidium elegans* (Chen et al., 2016)

Chemically-induced Deconstructing Strategy

TEMs from a dilute suspension of CNC from:

- (ae) rice straw (Lu and Hsieh, 2012)
- (af) sisal (Siqueira et al., 2009)
- (ag) sugar beet pulp (Azizi Samir et al., 2004)
- (ah) soy hulls (Flauzino Neto et al., 2016)
- (ai) tomato peel (Jiang and Hsieh, 2015)
- (aj) tunicin (Anglès and Dufresne, 2000)
- (ak) waste newspaper (Danial et al., 2015)
- (al) waste sackcloth (Cao et al., 2015)
- (am) wheat straw (Helbert et al., 1996)
- (an) wood fiberboard waste (Courret et al., 2017)



The Need for International Standards - Terminology

| Acronym | Terminology | Reference |
|---------|----------------------------------|--|
| – | Cellulose Micelles | (Rånby, 1949) |
| – | Level-off D.P. Cellulose Product | (Battista and Smith, 1961) |
| Wh | Whiskers | (Helbert et al., 1996; Dufresne, 2008) |
| – | Cellulose Crystallites | (Dong et al., 1996) |
| – | Cellulose Microcrystals | (Araki et al., 2001) |
| CNC | Cellulose Nanocrystals | (Grunert and Winter, 2002; Paralikar et al., 2008; Mangalam et al., 2009) |
| CNF | Cellulose Nanowhiskers | (Oksman et al., 2006; Petersson et al., 2007; Habibi et al., 2008; Braun et al., 2008; Rojas et al., 2009) |
| – | Nanocellulose | (Morán et al., 2008) |
| NCC | Nanocrystalline Cellulose | (Bai et al., 2009) |

TAPPI, ISO and
CSA approved

Table 3.1: Different terminologies used in the literature to describe the material resulting from the cellulose fiber fibrillation process.

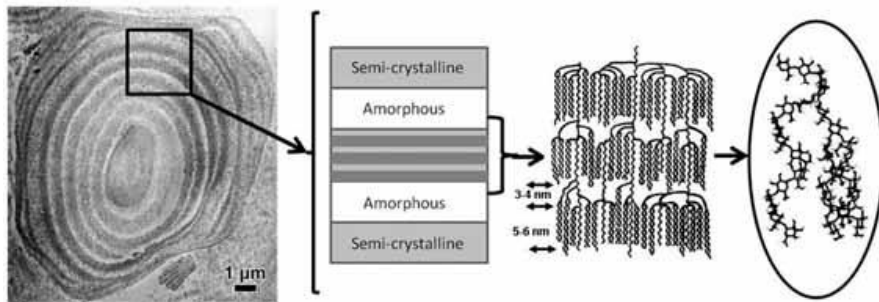
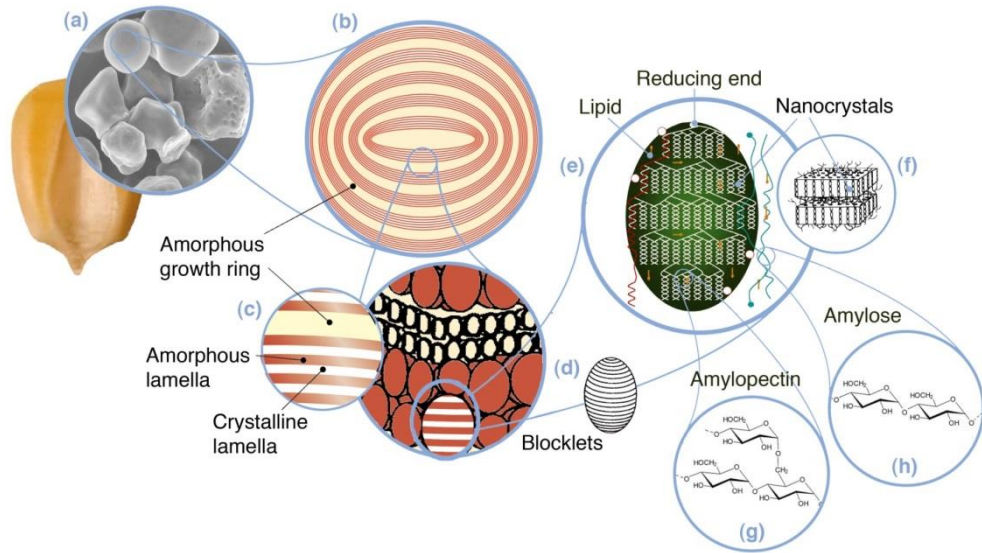
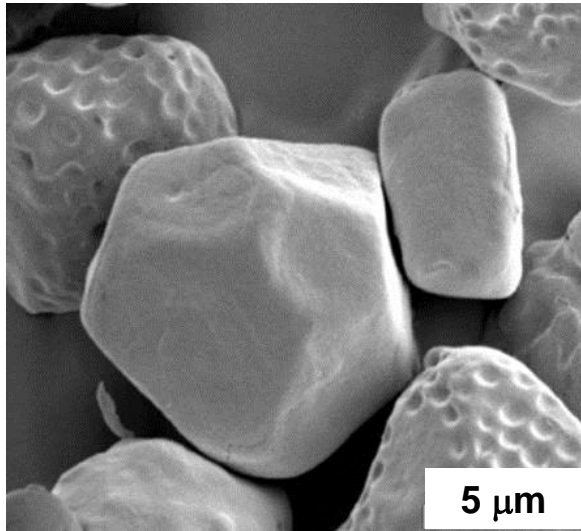
Starch

Starch = storage polymer → native starch = discrete and partially crystalline microscopic granules

corn, wheat, rice,
potato, tapioca, peas

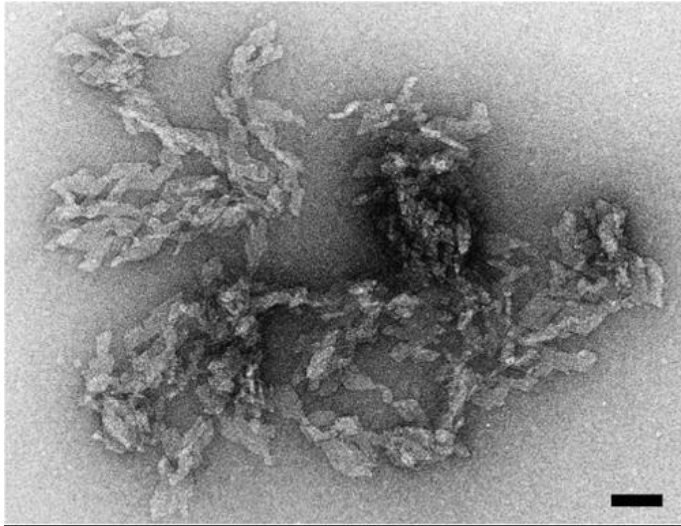


Starch



- (a) starch granules
- (b) amorphous and semi-crystalline growth rings
- (c) amorphous and crystalline lamellae
- (d) blocklets, (f) nanocrystals
- (g) amylopectin, (h) amylose

Starch Nanocrystals

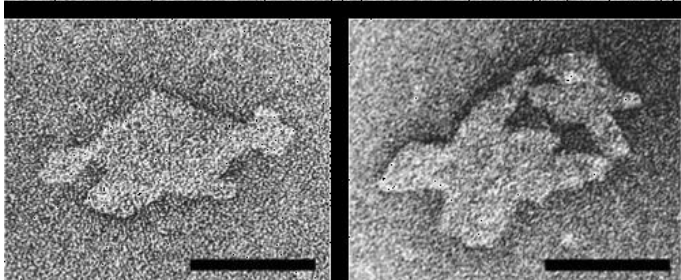
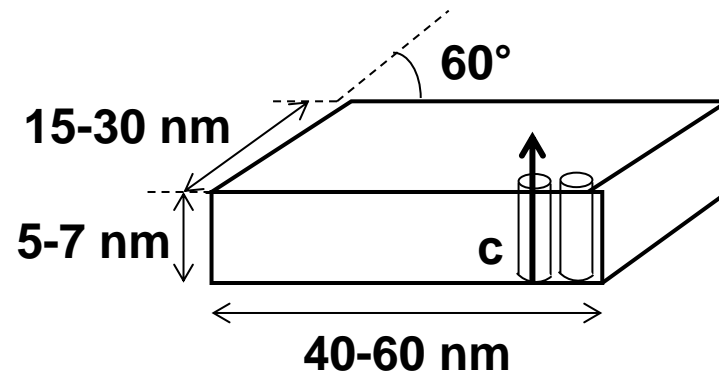
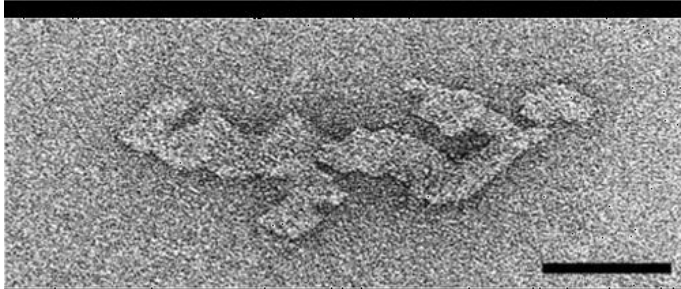


TEM of negatively stained SNC obtained after 3.16 M H_2SO_4 hydrolysis of waxy maize starch granules during 5 days, at 40 °C, 100 rpm and with a starch concentration of 14.69 wt %

(a) Aggregates of nanocrystals

(b-d) organizations of nanoplatelets

Scale bar: 50 nm.



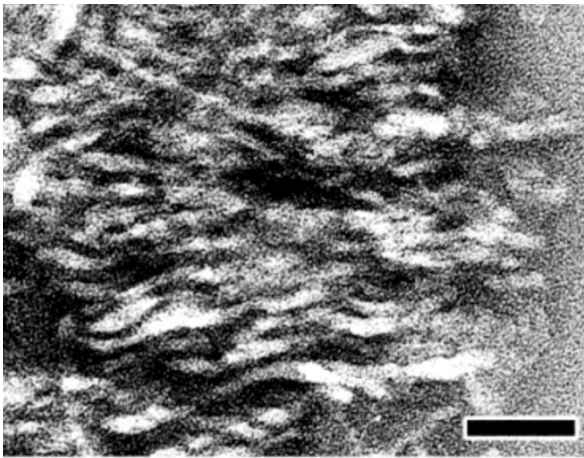
Putaux et al., *Biomacromolecules*
2003, 4, 1198-1202



Angellier et al., *Biomacromolecules*
2004, 5, 1545-1551

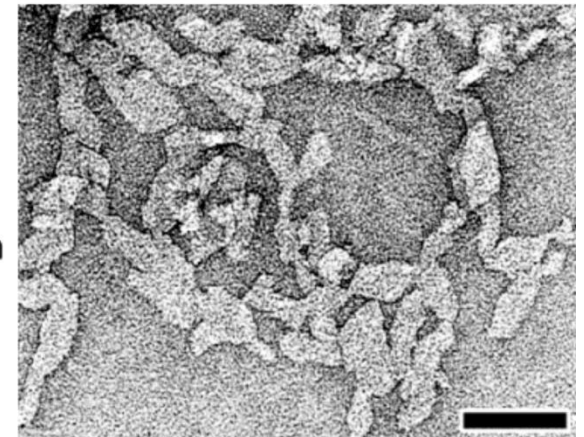
Starch Nanocrystals

2 weeks hydrolysis (2.2 N HCl at 36°C)

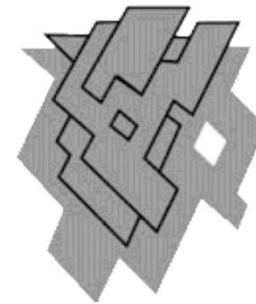
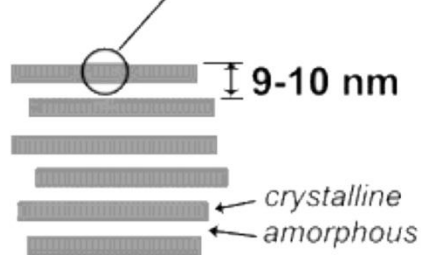
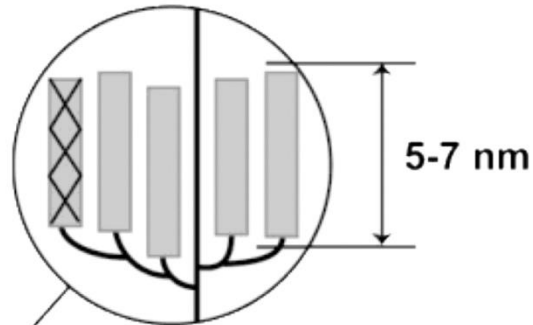


Longitudinal view

6 weeks hydrolysis (2.2 N HCl at 36°C)



Planar view



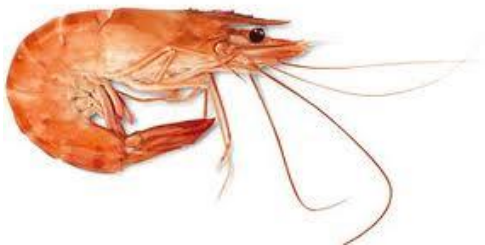
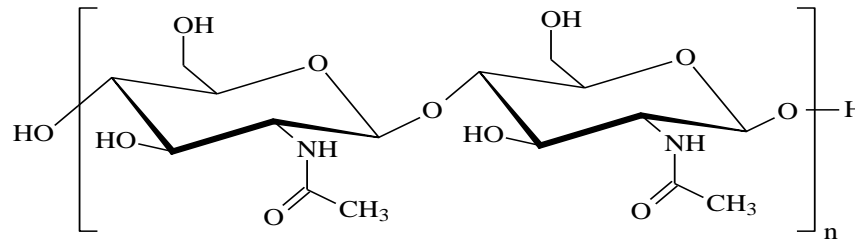
<https://www.youtube.com/watch?v=hdx2A5gq9js>

LGP²

Putaux et al., *Biomacromolecules*
2003, 4, 1198-1202

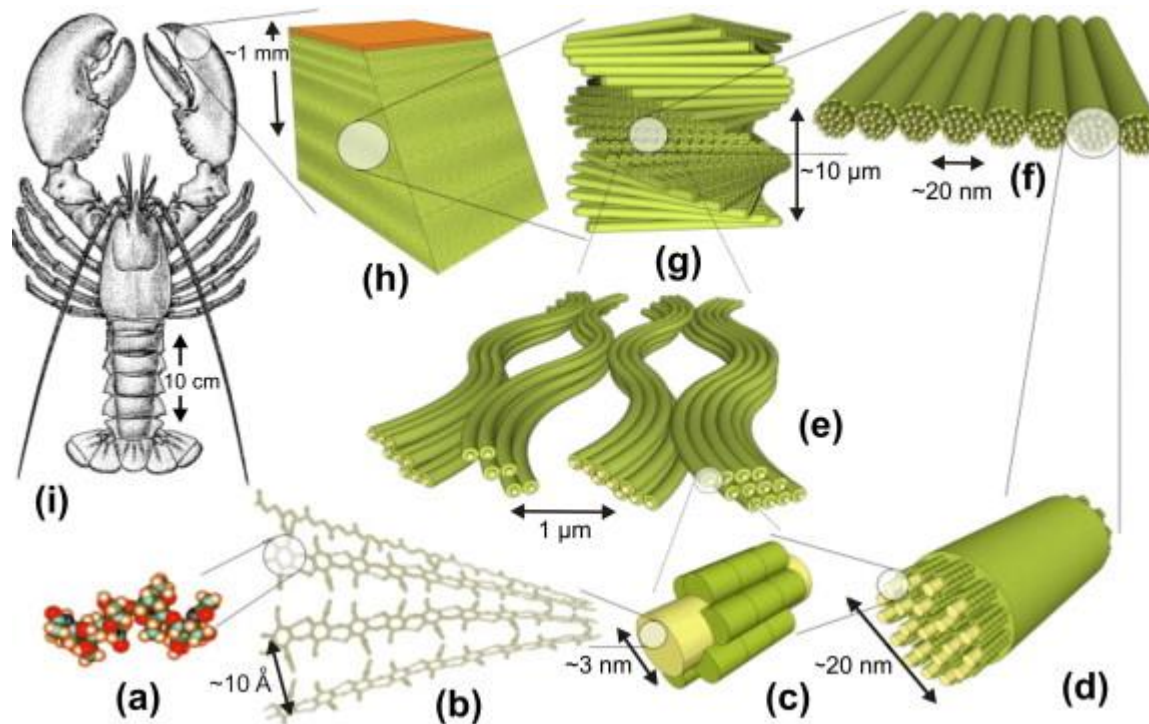
Chitin

Main component of the cell walls of fungi, the exoskeletons of arthropods (crabs, lobsters, shrimps) and insects, the radulas of mollusks, and the beaks of cephalopods (squid, octopuses)



Chitin

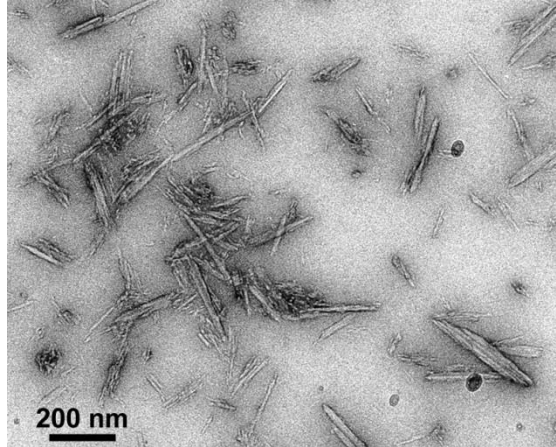
Exoskeleton of lobster (*Homarus americanus*)



Nikolov et al., *J. Mech. Behav. Biomed.* 2011, 4, 129-145

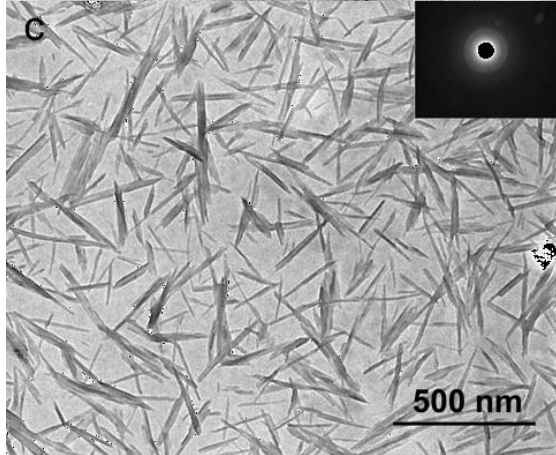
Chitin Nanocrystals

Squid pen

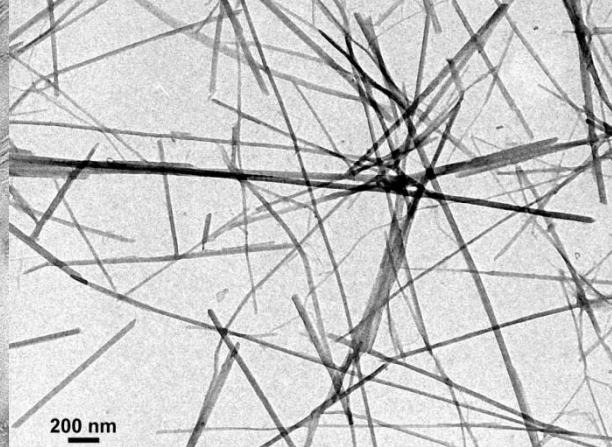


Paillet and Dufresne,
Macromolecules **2001**,
34, 6527-6530

Crab shell



Gopalan Nair and
Dufresne,
Biomacromolecules
2003, 4, 657-665

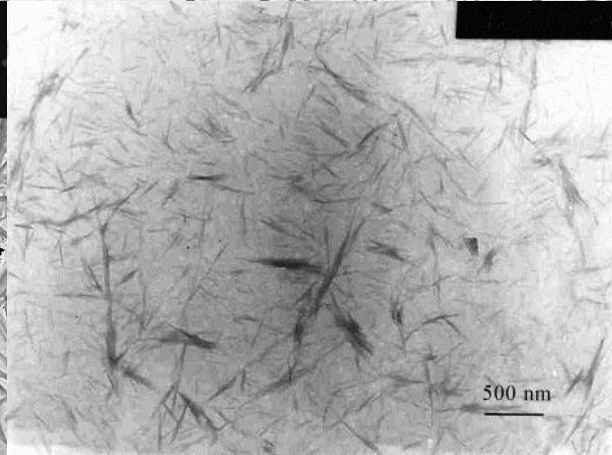


Riftia tubes



Morin and Dufresne,
Macromolecules **2001**,
35, 2190-2199

Shrimp shell



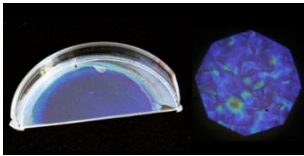
Sriupayo et al.,
Polymer **2005**, 46,
5637-5644

Applications of Cellulose Nanomaterials

Coatings



Films



Biomedical



Detergents



Adhesives



Energy



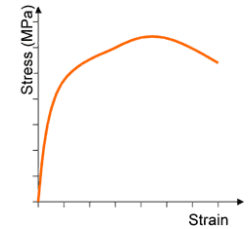
Inks/printing



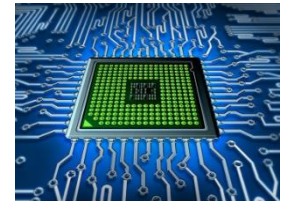
Pulp & paper



Composites



Electronics



Filtration



Textiles



Cosmetics



Food industry



Packaging

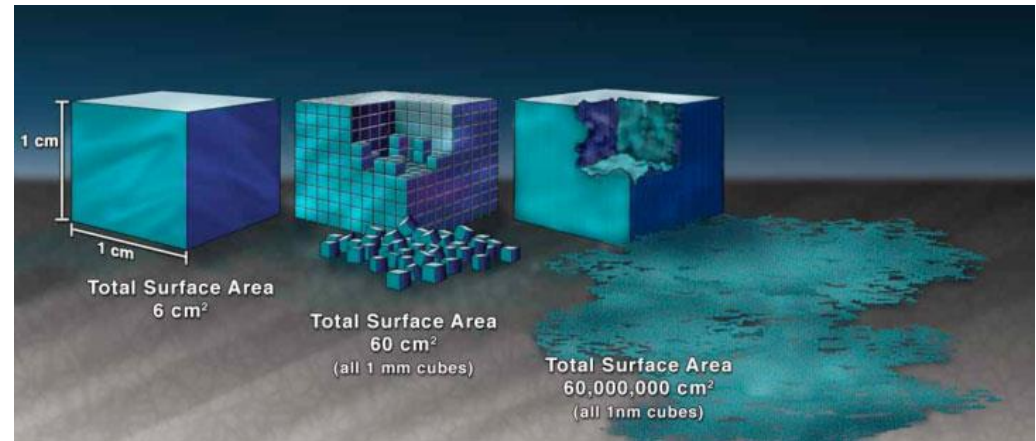
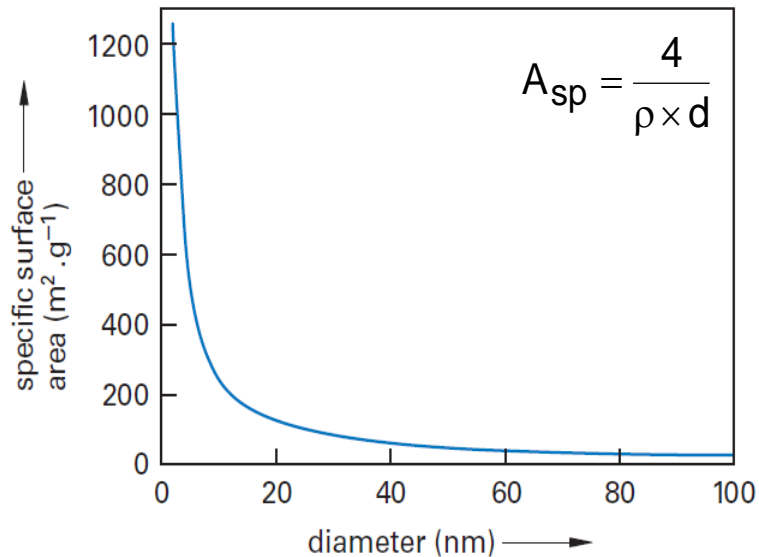


Construction



Nanoparticles vs. Microparticles

- ① Increase of the specific area ($\sim 100 \text{ m}^2 \cdot \text{g}^{-1}$ vs. $\sim 1 \text{ m}^2 \cdot \text{g}^{-1}$)



More surface is better surface !

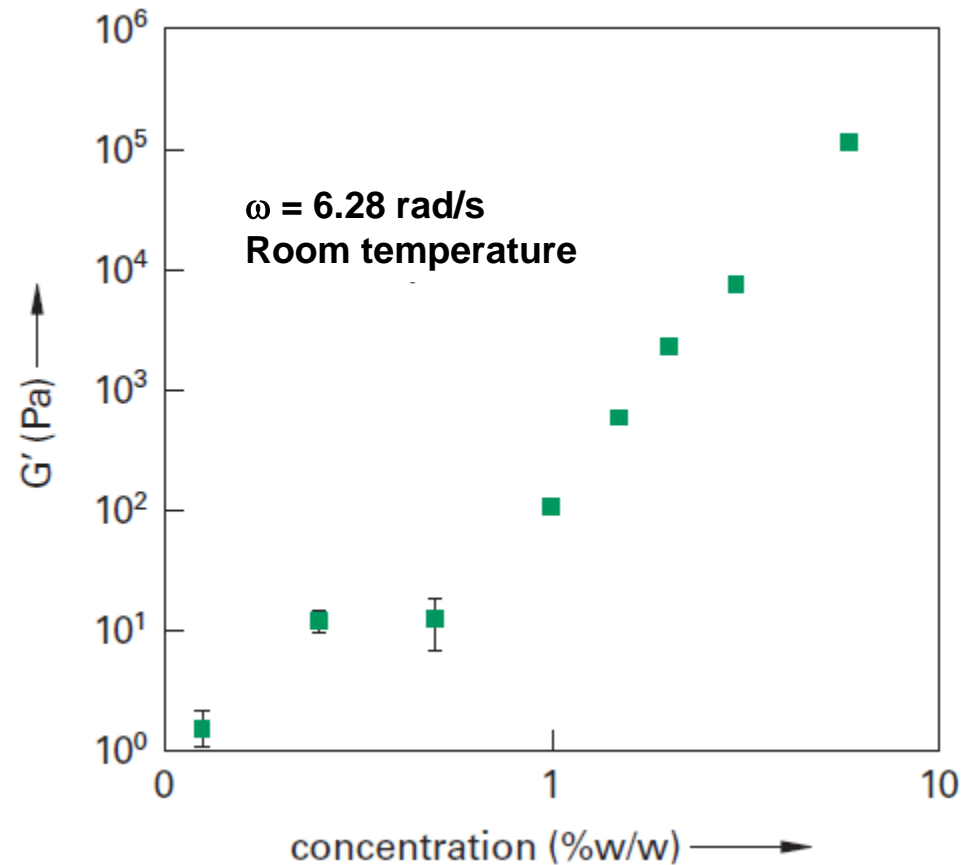
High Specific Surface Area



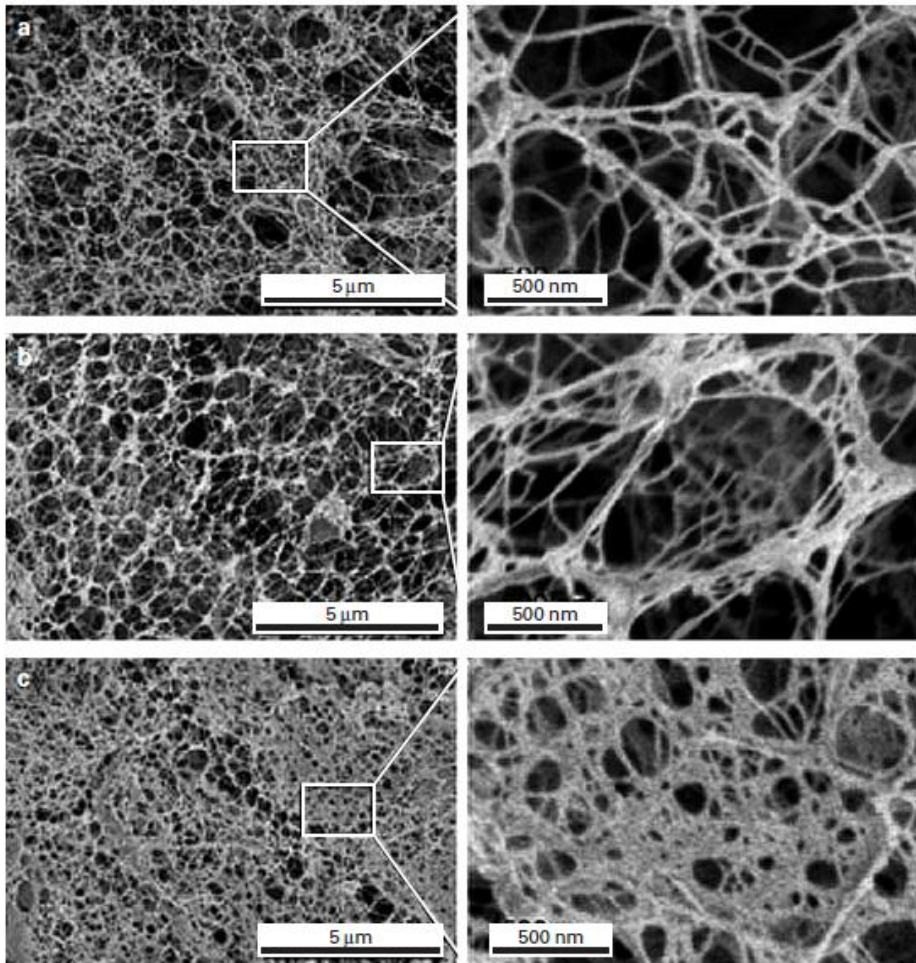
Lavoine et al., *Carbohydr. Polym.*
2012, 90, 735-764

Applications

Food, cosmetic, pharmaceutical
industries



High Specific Surface Area



Production of foams and aerogels

Density

(a) 7 kg.m^{-3}

(b) 32 kg.m^{-3}

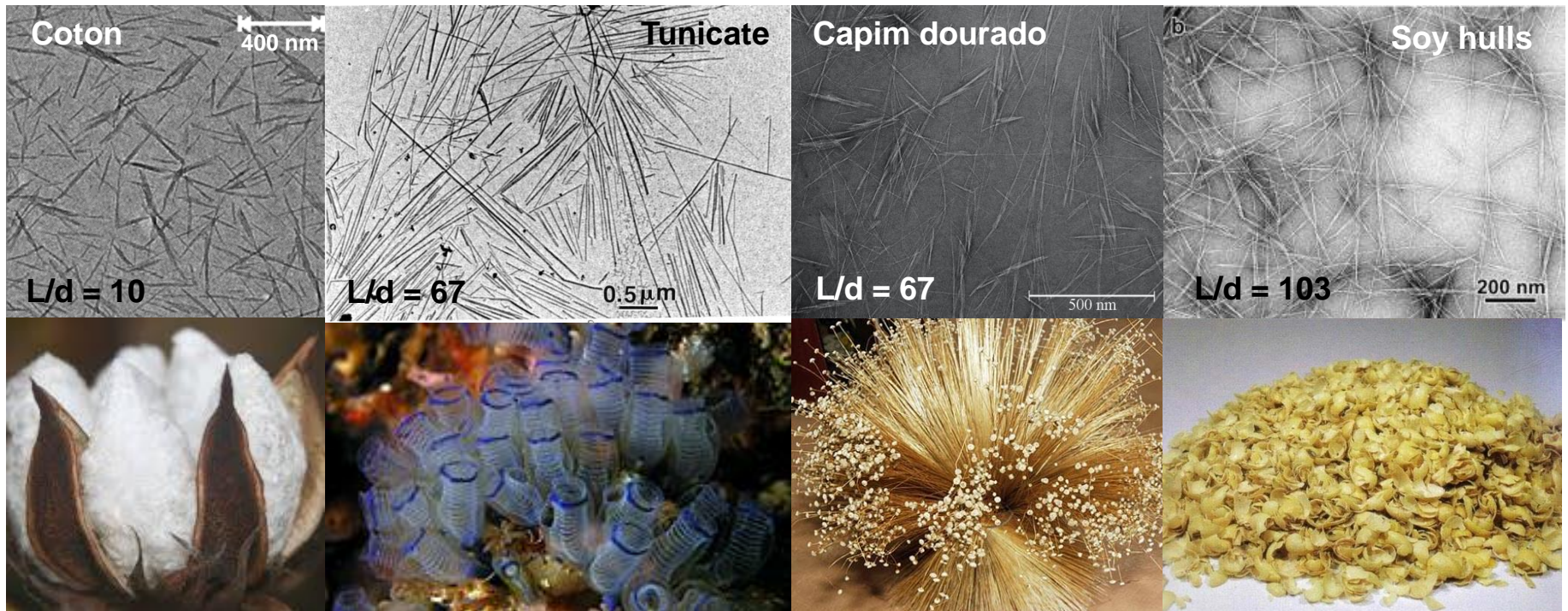
(c) 79 kg.m^{-3}

Applications

Porous templates, filtration

Nanoparticles vs. Microparticles

② High aspect ratio (10-100 for CNC, much higher for CNF)



Fleming et al., *J. Am. Chem. Soc.* **2010**, 122, 5224-5225

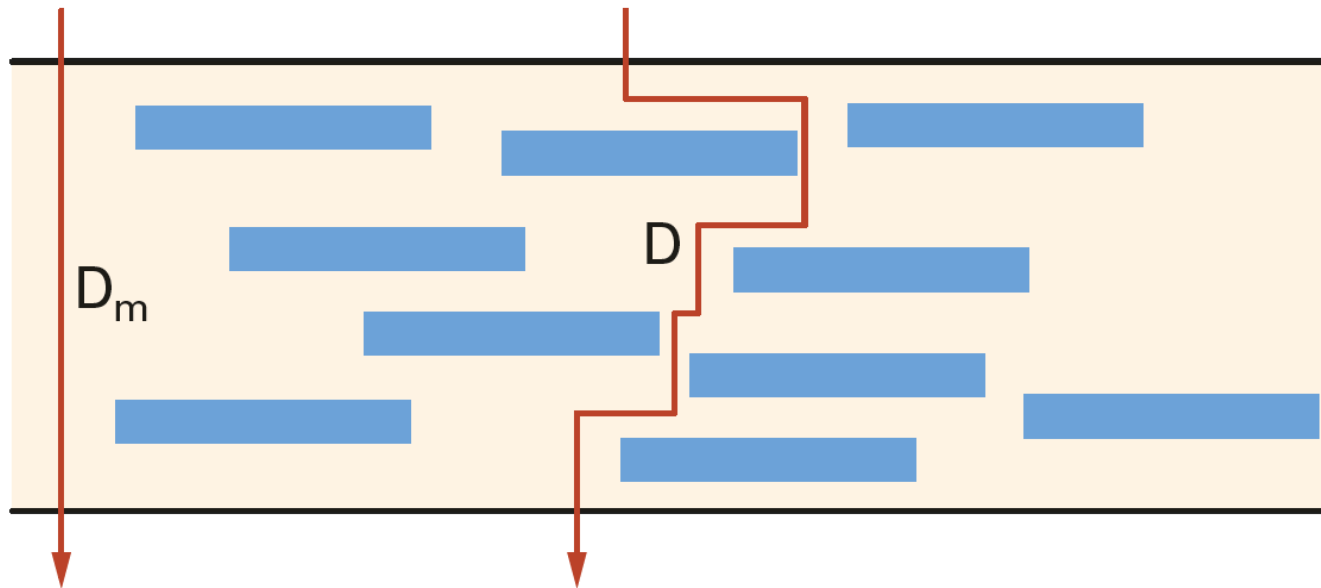
Anglès and Dufresne, *Macromolecules* **2000**, 33, 8344-8353

Siqueira et al., *Cellulose* **2010**, 17, 289-298

Flauzino Neto et al., *Carbohydr. Polym.* **2016**, 153, 143-152

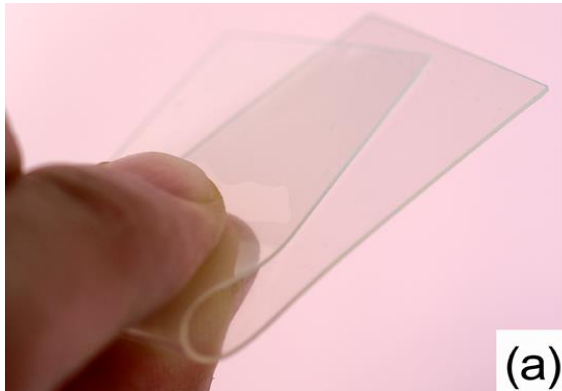
Nanoparticles vs. Microparticles

- ③ The average inter-particles distance decreases as their size decreases
→ particle-particle interactions
- ④ Nanoparticles are weight efficient: improved properties for low filler content without detrimental effect on impact resistance and plastic deformation
- ⑤ Reduction of gas diffusion (barrier effect)



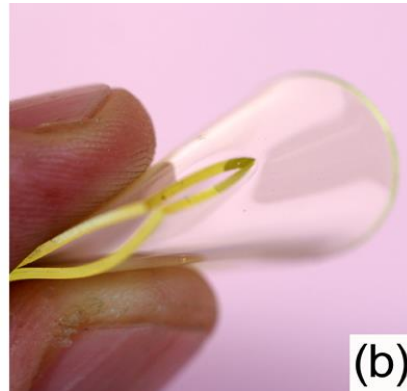
Nanoparticles vs. Microparticles

- ⑥ Small particles are "invisible": transparent coatings/films are attainable



Foldable transparent acrylic resin sheet with 5 wt% BC nanofibers

Nogi and Yano, *Adv. Mater.*, **2008**, 20, 1849-1852



More fragile neat acrylic resin sheet



Flexibility and transparency of acrylic resin film with 60 wt% BC nanofibers

Yano et al., *Adv. Mater.*, **2005**, 17, 153-155

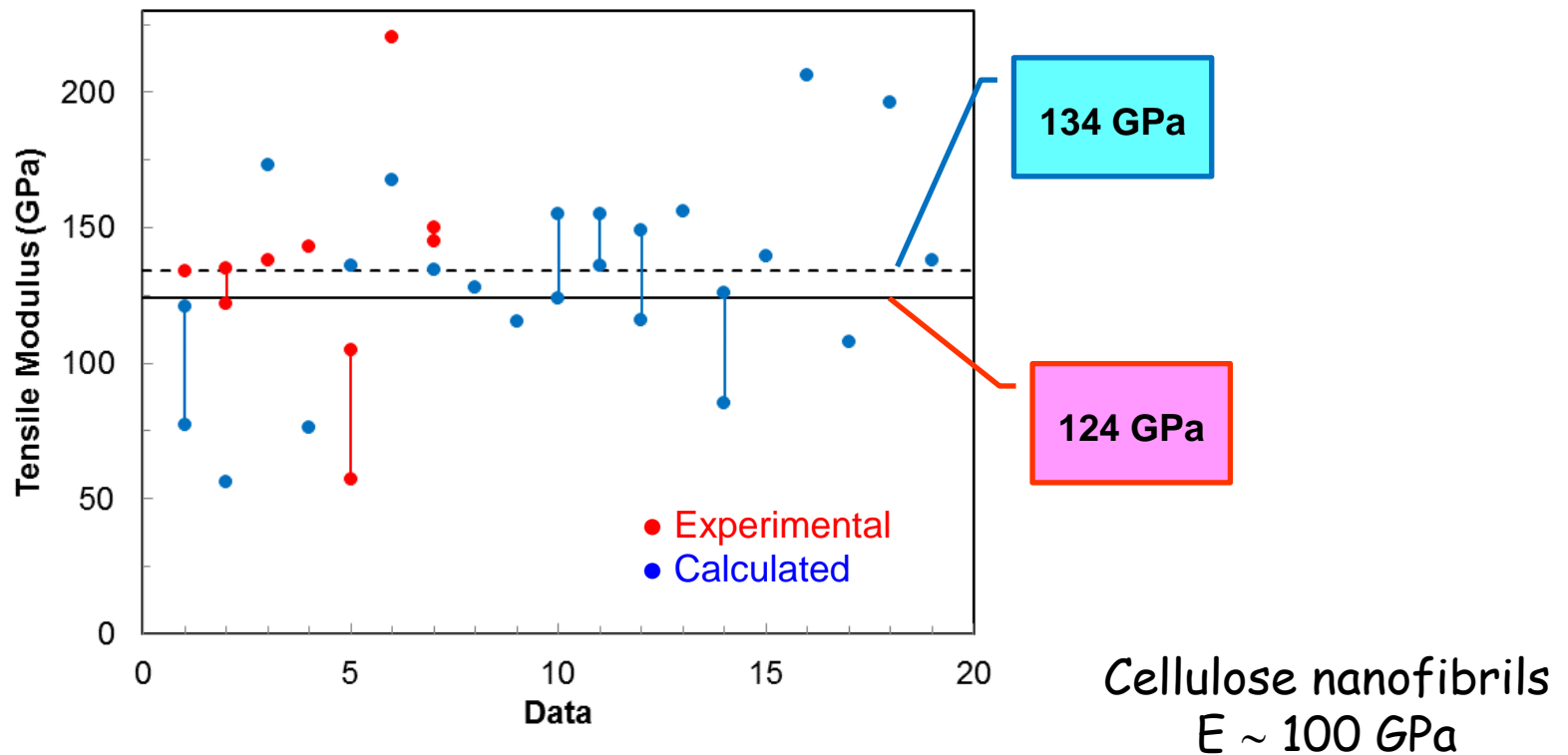
Applications

Electronics (flexible circuits)

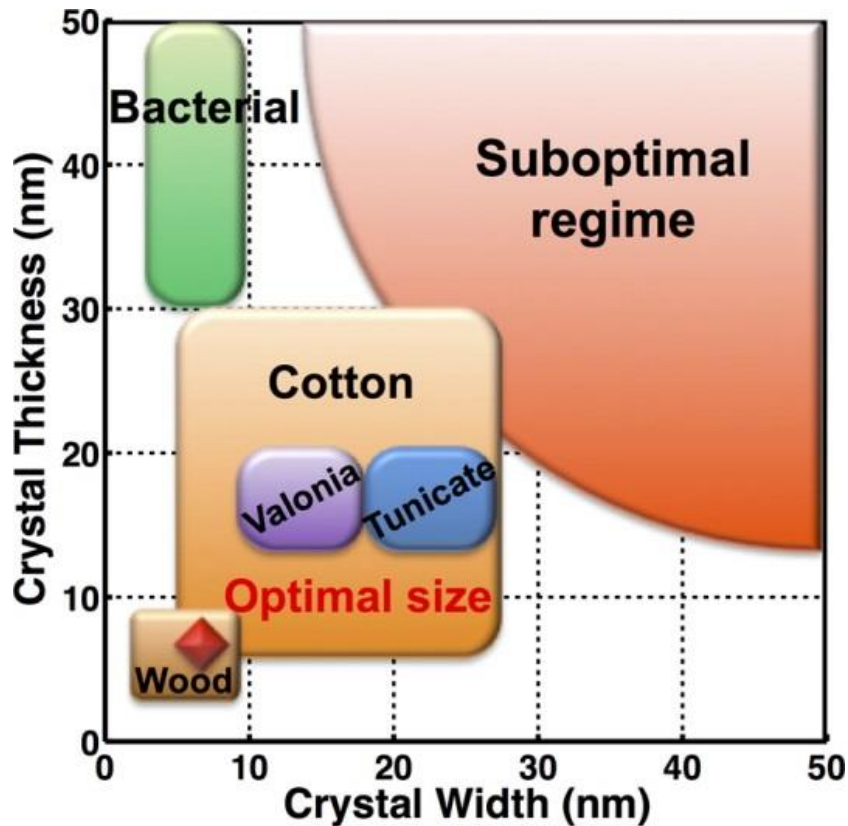
Energy (solar panels)

Nanoparticles vs. Microparticles

- ⑦ Because cellulose nanomaterials contain only a small number of defects, their axial Young's modulus is close to the one derived from theoretical chemistry



Nanoparticles vs. Microparticles



Atomistic simulations on the fracture energy of I β cellulose nanocrystals

Ideal dimensions optimizing fracture energy are:

4.8-5.6 nm in thickness (6-7 chain layers)

6.2-7.3 nm in width (6-7 chain layers)

Sinko et al., *ACS Macro Lett.* **2014**, 3, 64-69

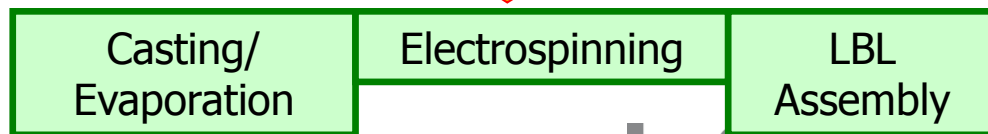
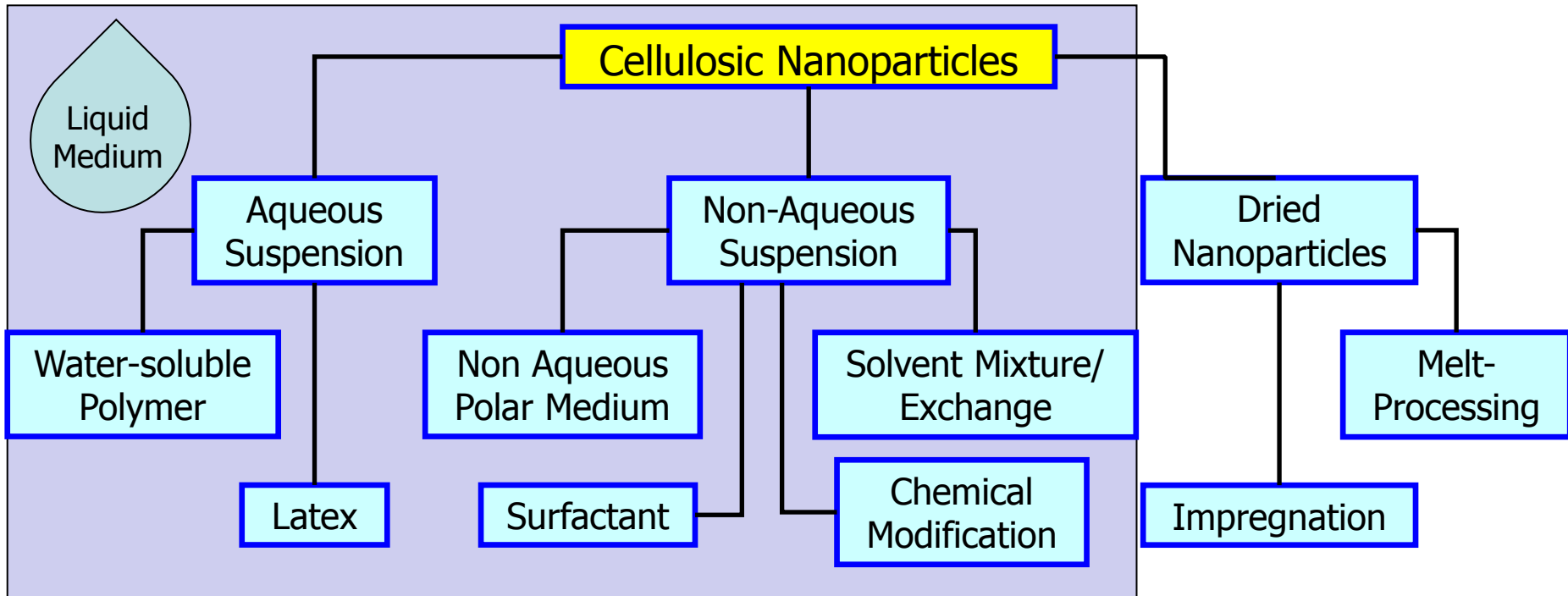
Nanoparticles vs. Microparticles

⑧ **Lightweight material:** Cellulose nanomaterial modulus potentially stronger than steel and similar to Kevlar

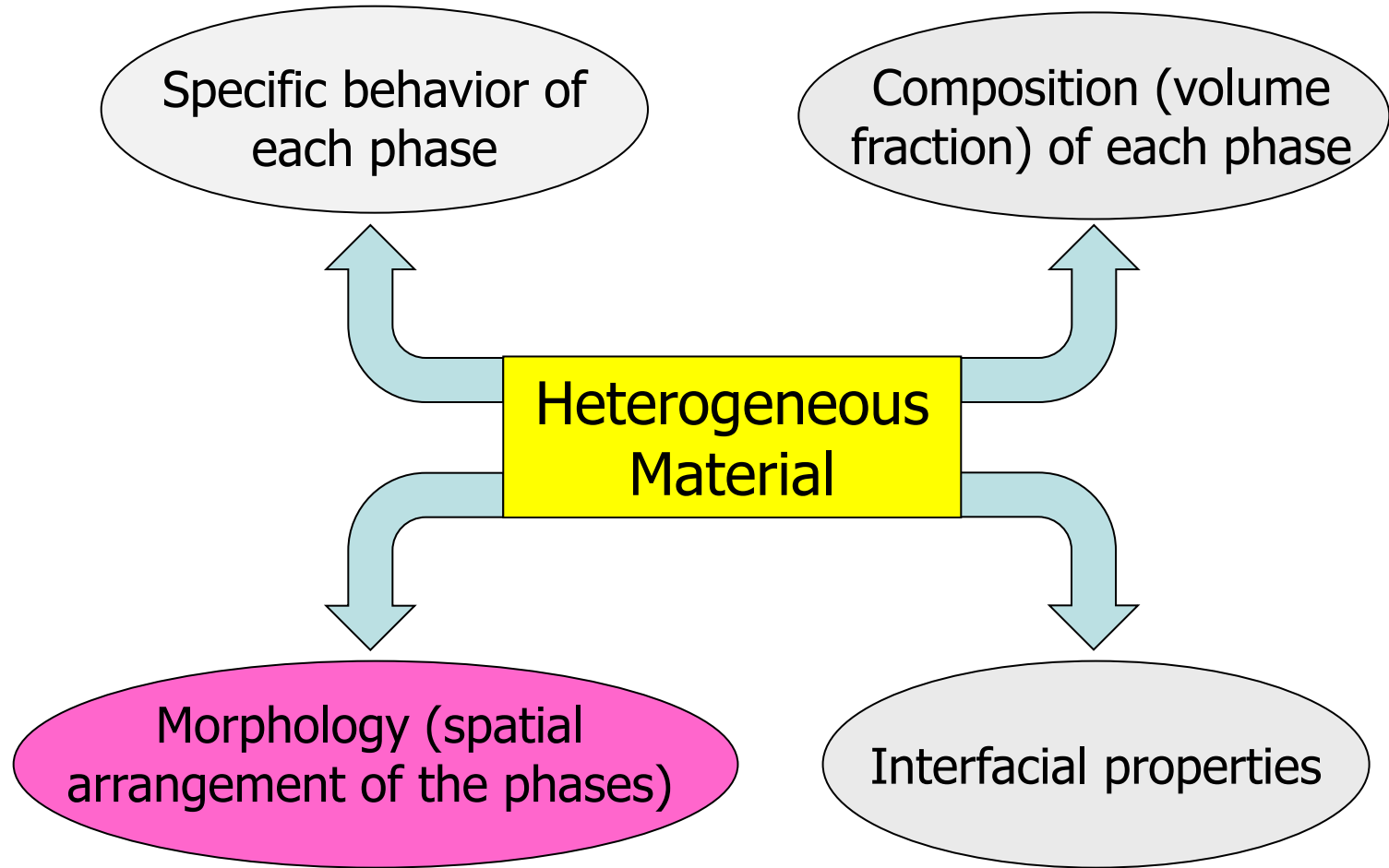


| Material | Modulus (Gpa) | Density (g.cm^{-3}) | Specific Modulus (J.g^{-1}) |
|----------|---------------|--------------------------------|--|
| Glass | 70 | 2.6 | 27 |
| Kevlar | 60-125 | 1.45 | 41-86 |
| Steel | 200-220 | 8 | 25 |
| MFC | 100 | 1.5-1.6 | 65 |
| CNC | 130 | 1.5-1.6 | 85 |

Processing of Nanocomposites



Cellulose Based Nanocomposites

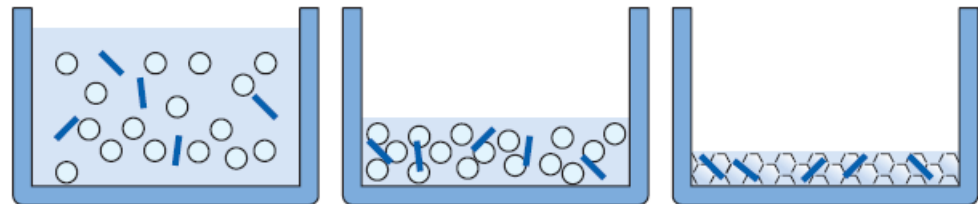
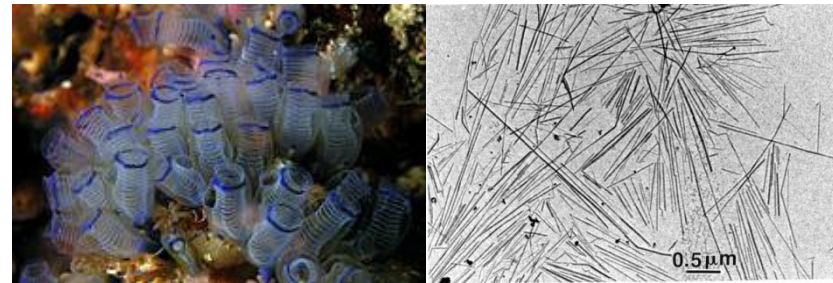
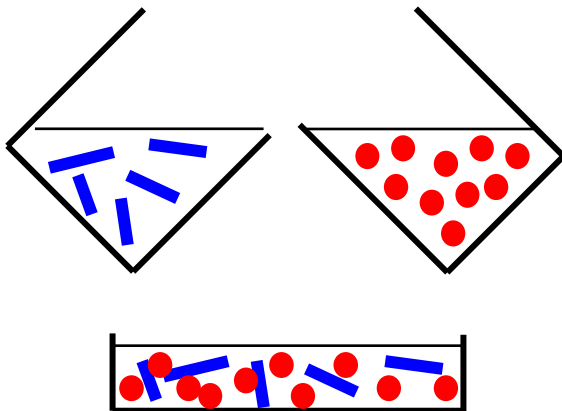


Cellulose Nanocomposites – Pioneering Work

Preferred processing medium = water because of high stability of aqueous cellulose nanomaterial dispersions

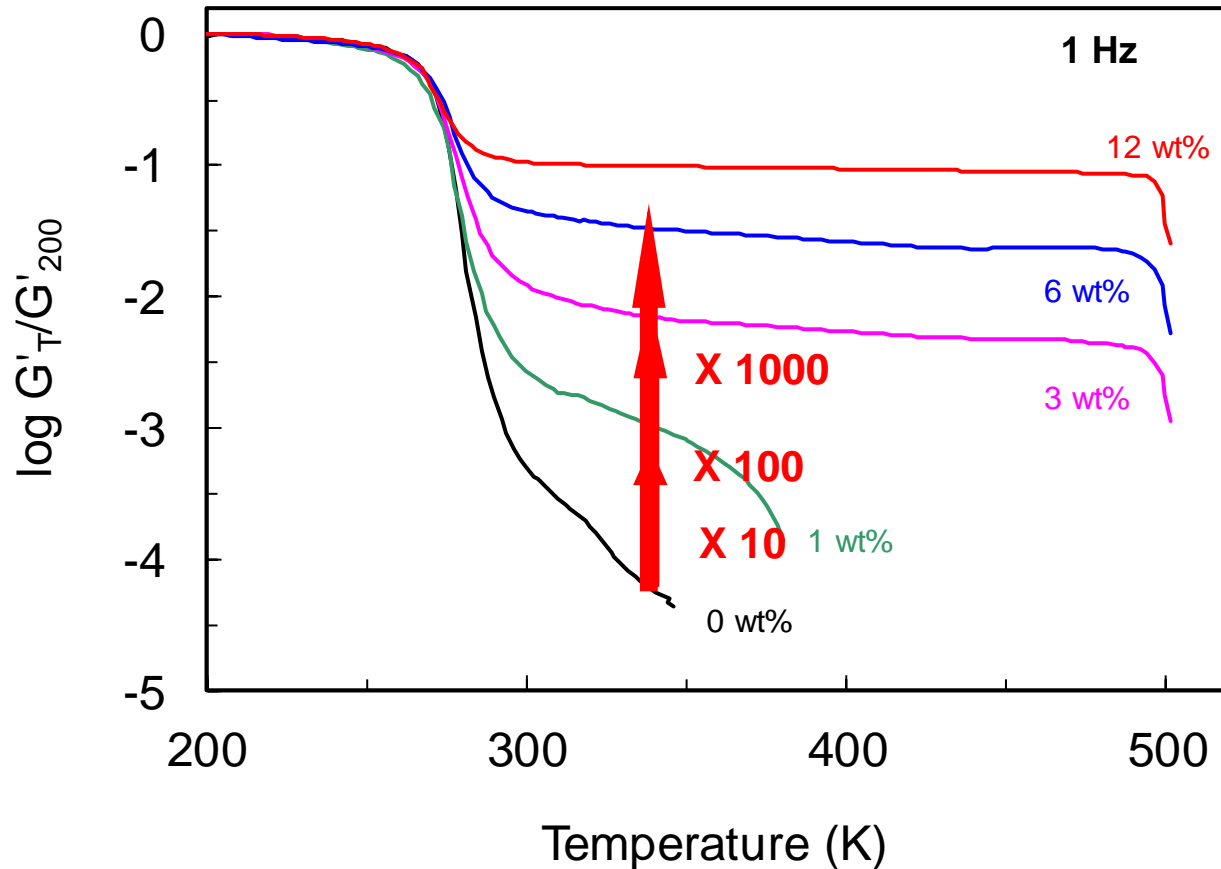
Matrix = water-soluble polymer or latex (poly(S-co-BuA))

water evaporation ($T > T_g$) → particle coalescence → nanocomposite film



Favier et al., *Polym. Adv. Technol.*, **1995**, 6, 351-355

Cellulose Nanocomposites – Pioneering Work

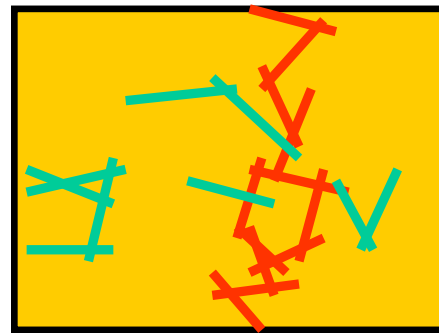
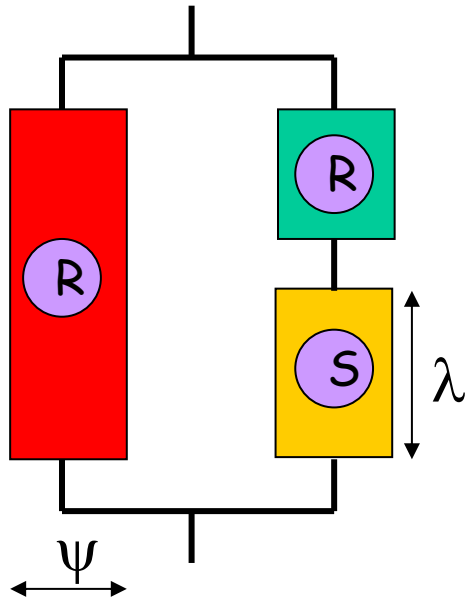


High reinforcing effect at $T > T_g$

Thermal stabilization up to 500 K ($\phi_R > 1\text{wt}\%$) (degradation cellulose)

Cellulose Nanocomposites – Pioneering Work

Percolation Approach: Takayanagi Model



percolating whiskers network :
 $E_R = 15 \text{ GPa} \rightarrow G_R = 5 \text{ GPa}$

$$G = \frac{(1 - 2\psi + \psi\phi_R)G_S G_R + (1 - \phi_R)\psi G_R^2}{(1 - \phi_R)G_R + (\nu_R - \psi)G_S}$$

if $G_R \gg G_S \Rightarrow G = \psi G_R$

$$\psi = 0 \quad \text{for } \phi_R < \phi_{Rc}$$

$$\psi = \phi_R \left(\frac{\phi_R - \phi_{Rc}}{1 - \phi_{Rc}} \right)^b \quad \text{for } \phi_R \geq \phi_{Rc}$$

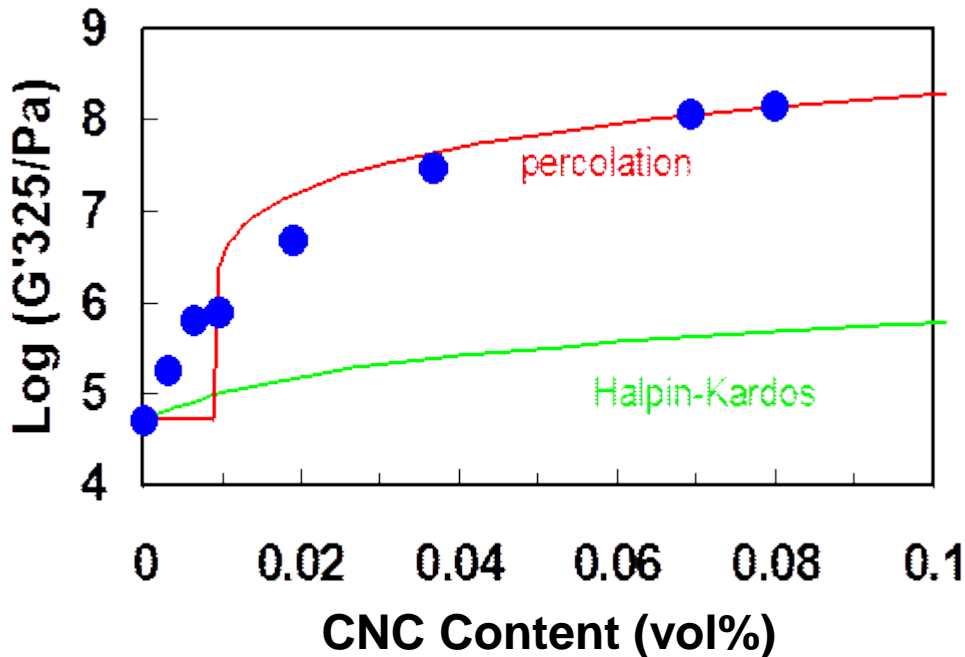
$$L/d = 67 \rightarrow \phi_{Rc} = 1\%$$

$$b = 0.4 \text{ (3D system)}$$

- ψ = volume fraction of the percolating rigid phase
- ϕ_R = volume fraction of filler
- ϕ_{Rc} = critical volume fraction at the percolation threshold
- b = critical exponent
- G_R = modulus of the percolating CNC network

Favier et al., *Polym. Adv. Technol.*, **1995**, 6, 351-355

Cellulose Nanocomposites – Pioneering Work



Good agreement between experimental and predicted data



Strong interactions between CNCs (H-bonding forces)
→ formation of a rigid cellulose CNC network for $\phi_R > \phi_{RC}$

Mechanical percolation effect

- High reinforcing effect
- Thermal stabilization of the composite modulus
(water evaporation = slow process)

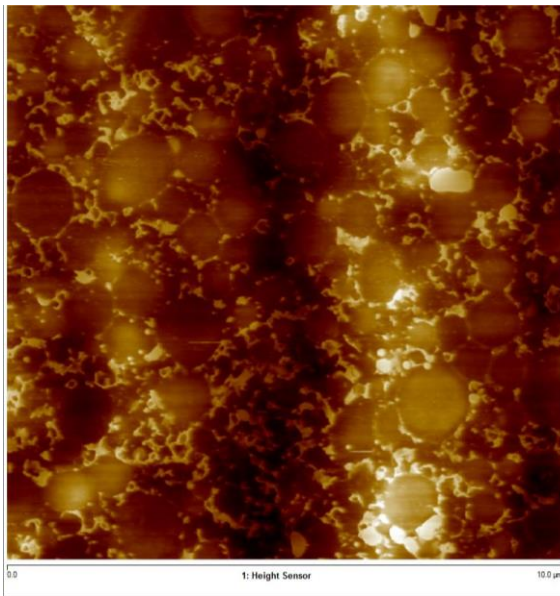
Favier et al., *Polym. Adv. Technol.*, 1995, 6, 351-355

Percolation Threshold

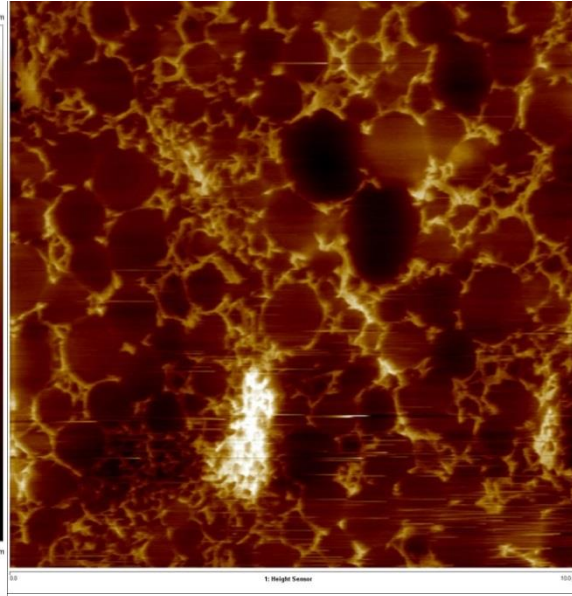
| Source | CNC | | | |
|------------------|--------|--------|-----|-----------------|
| | L (nm) | D (nm) | L/D | Φ_R (vol%) |
| Cotton | 170 | 15 | 10 | 7 |
| Flax | 300 | 20 | 15 | 4.6 |
| Sisal | 250 | 4 | 60 | 1.1 |
| Luffa | 183 | 5 | 37 | 1.8 |
| Sugar beet Pulp | 210 | 5 | 42 | 1.3 |
| Palm tree rachis | 260 | 6 | 43 | 1.3 |
| Palm tree foliol | 180 | 6 | 30 | 2.3 |
| Wheat straw | 220 | 5 | 45 | 1.6 |
| Hard wood | 200 | 4 | 50 | 1.4 |
| Soft wood | 200 | 4 | 50 | 1.4 |

$$\Phi_R = \frac{0.7}{L/D}$$

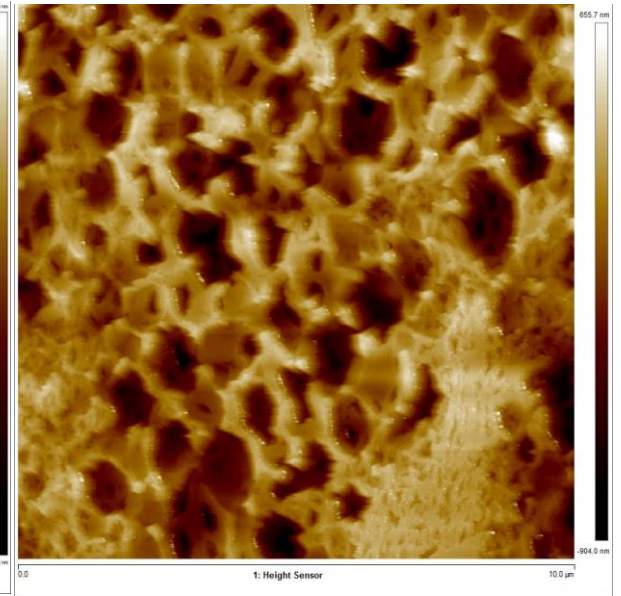
Percolation Network



NR



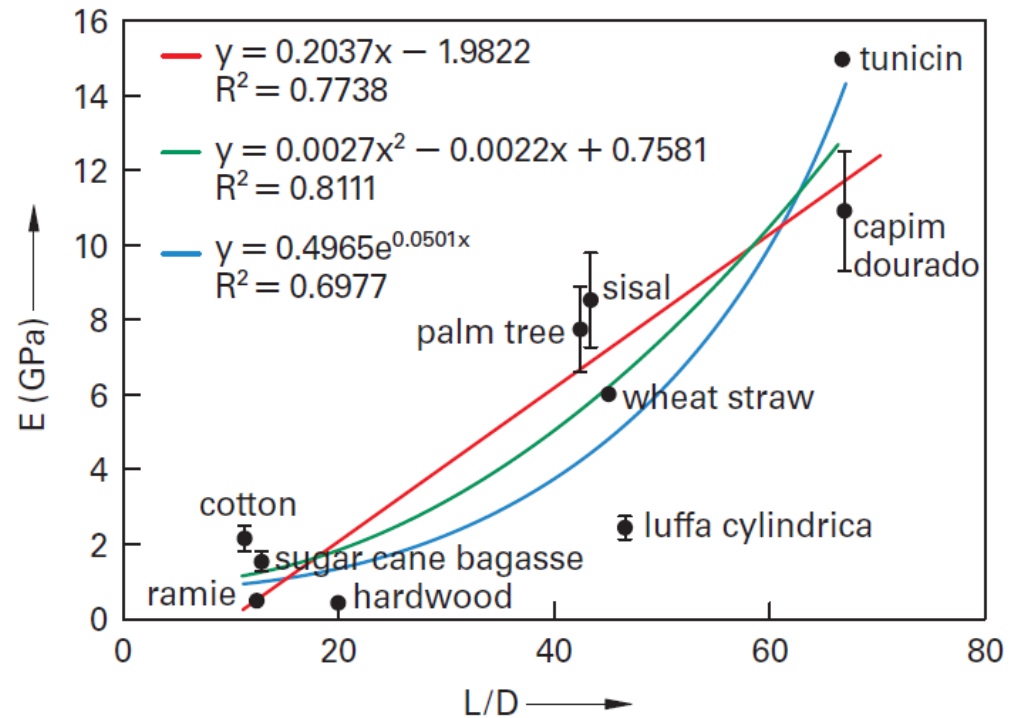
NR + 8.2 wt% CNC



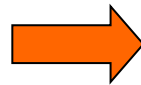
NR + 16.4 wt% CNC

Stiffness of the Percolating Network

$$G = \psi G_R$$



High L/d CNC



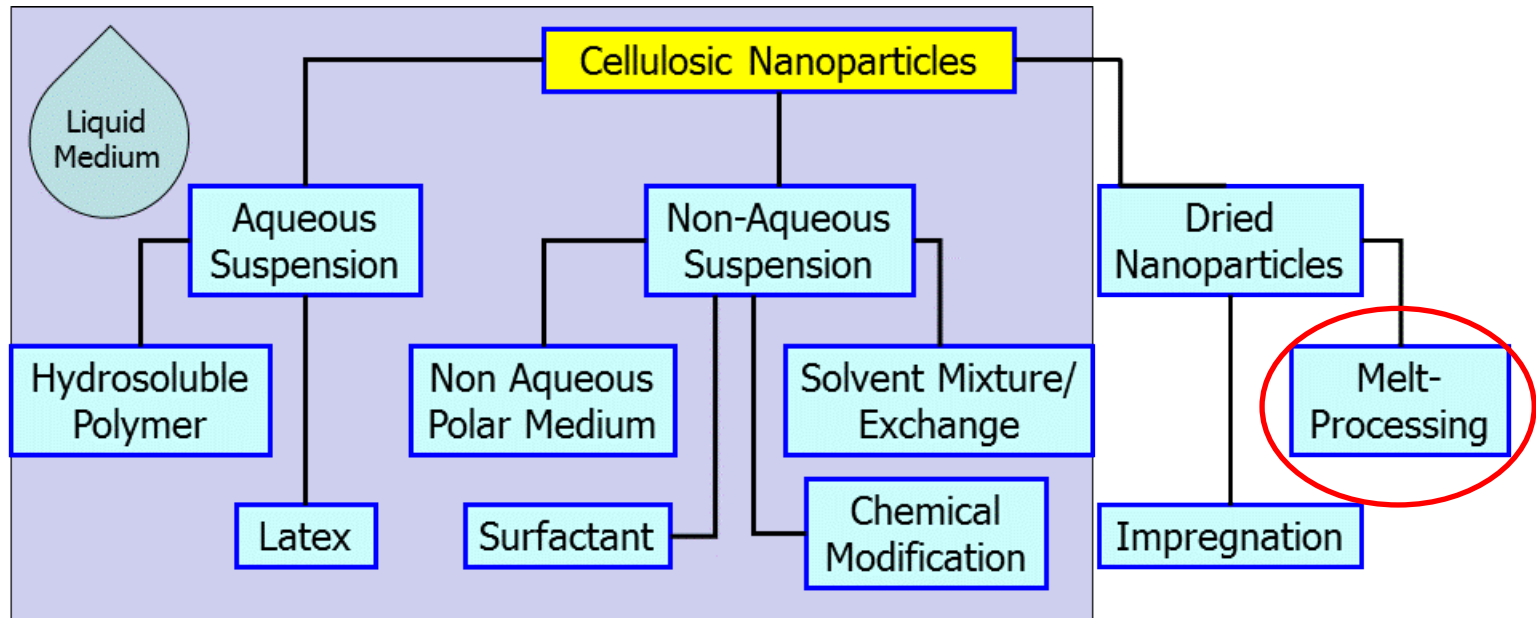
low percolation threshold

High stiffness of the percolating CNC network

Bras et al., *Carbohydr. Polym.* 2011, 84, 211-215



Processing of Nanocomposites



Highly challenging !

Processing of Nanocomposites

Solvent/wet approach (casting/evaporation)

Preservation of the dispersion state in the liquid medium

Limitation of the number of polymer matrices

Non-industrial and non-economic

Polymer melt approach (extrusion, injection molding)

Green process

Industrially and economically viable

Hydrophilicity → aggregation of cellulosic nanoparticles upon drying

Difficulties for uniform dispersion within the polymer melt

Low thermal stability

Structural integrity of the nanoparticle

Orientation of the nanoparticle

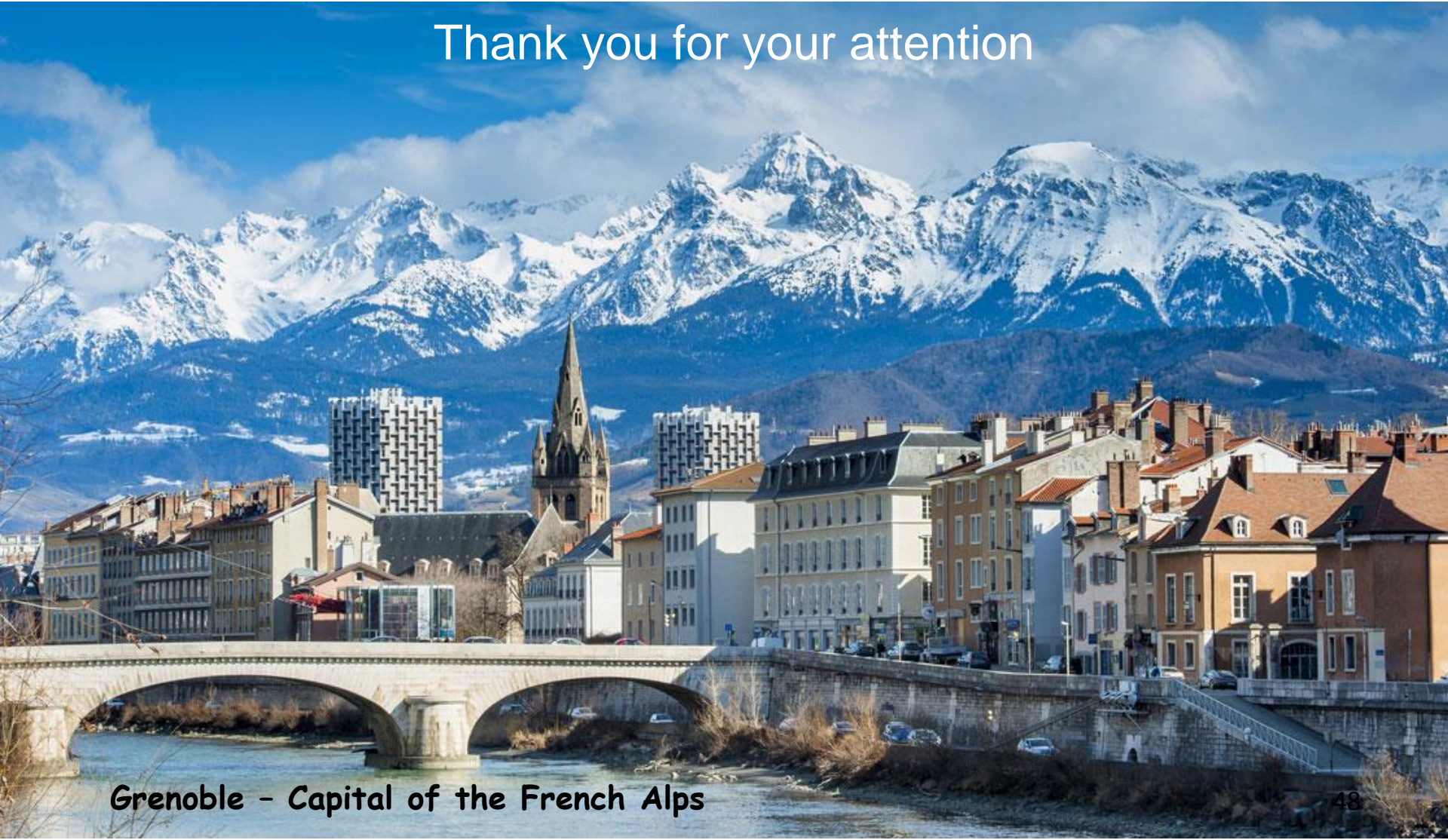
Conclusion

- ➔ Growing interest in both the non-food usage of renewable resources and nanosized particles
- ➔ Polysaccharide : low cost material, abundant, renewable
- ➔ Preparation of nanoparticles with different aspect ratios
- ➔ Nanosized particles : mechanical properties (strength, modulus, dimensional stability), decreased permeability to gases and water, thermal stability, heat distortion temperature
- ➔ Many possible applications: optical, mechanical, barrier, rheological properties
 - Sustainability of supply
- ➔ **Challenges:** Melt processing of cellulose based nanocomposites
 - Improvement of nanocomposite properties in moist atmosphere



agefpi

Thank you for your attention



Grenoble - Capital of the French Alps